STUDY REGARDING THE HONEY CONTAMINATION DEGREE ASSESSED IN A SPECIALIZED PRODUCTION UNIT

Carmen Daniela PETCU, Emilia CIOBOTARU-PÎRVU, Oana-Mărgărita GHIMPEȚEANU, Gheorghe Valentin GORAN, Corina Nicoleta PREDESCU, Oana Diana OPREA

University of Agronomic Sciences and Veterinary Medicine of Bucharest, Faculty of Veterinary Medicine, 105 Splaiul Independenței, District 5, 050097, Bucharest, Romania

Corresponding author email: carmen28petcu@gmail.com

Abstract

Honey represents a pleasant, nourishing food, with great biological and calorical value (315 kcal/100 g), easily digestible. It possesses real bactericidal properties, due to inhibin content. Examination of honey is necessary in order to assess its quality and purity, as well as to identify forbidden substances. The study was conducted in a processing unit which markets honey in the European Community. This study aimed to evaluate the contamination degree of the commercialised honey. In the summer of 2019, 3 batches of honey were analyzed (2 acacia honey batches containing 15 samples each and 1 batch containing 15 samples of polyfloral honey). Laboratory assessments were focused on determination of nitroimidazoles residues (metronidazole, dimetridazole, ronidazole), tetracyclines (cxytetracycline, tetracycline, chlortetracycline, doxycycline, demeclocycline, methacycline, minocycline), macrolides (clindamycin, erythromycin, josamycin, kitasamycin, lincomycin, oleandomycin, spiramycin, mirosamycin, tilmicosin, trimethoprim, tylosin), nitrofuran metabolites, chloramphenicol, streptomycin, dihydrostreptomycin, sulfonamides, trimethoprim, glyphosate. The methods used in the research were HPLC, LC-MS/MS, ELISA. The final results of the study that considered the a forementioned batches, sampled from various local beekeepers, proved that antimicrobial drug residues were in accordance with the national and international regulations, permitting marketing without restrictions.

Key words: antibiotics residues, chloramphenicol, honey, streptomycin.

INTRODUCTION

Honey represents the main product of beekeeping, being the result of nectar or manna bee processing and its storage in the honeycomb cells. Honey produced by bees exclusively from other raw materials than the one they naturally harvest, does not get into the frame of honey (Dolis, 2009). Honey is a natural food obtained in conventional or ecological systems, in units which follow the food safety principles, composed mainly of sugars and other constituents: enzymes, amino acids, organic acids, carotenoids, vitamins, minerals and aromatic substances (Petcu, 2006; Crivineanu et al., 2011; Tănăsoiu et al., 2014; Dobre, 2016; Dobre, 2017; Tăpăloagă et al., 2017; Tăpăloagă, 2018; Tudoreanu et al., 2012). It is rich in flavonoids and phenolic acids that act as natural antioxidants, being a beneficial product for consumers (Algarni et al., 2012; Tănăsoiu et al., 2015; Șapcaliu et al., 2017; Tamas-Krumpe, 2019).

In terms of food safety, honey has to be free of chemical, toxic and carcinogenic contaminants. especially pesticides and antibiotics (Orso et al., 2015; Murariu et al., 2019; Murariu et al., 2019). The most common and important contaminations of honey are done directly (treatments applied in the hive) and indirectly (contaminants that come from the agriculture and the environment) (Mărghitaș et al., 2010). Chemical contaminants have different origin: environmental pollutants (heavy metals), chemicals used in agriculture (pesticides), toxic contamination substances and those formed during processing and storage stages (disinfectants, detergents and mycotoxin, chemicals which could migrate from packagings or packaging systems), direct treatment of bees against bacterial diseases of the bee brood, like American foulbrood or European foulbrood (Petcu, 2104a; Petcu et al., 2014b; Tofană, 2011).

Sulfonamides, tetracyclines, nitrofurans and macrolides are used by beekepers for

preventing and controlling honeybee diseases. Consequently, it is possible that antibiotics residues from honey may be the result of treatments carried out by beekeepers. The treatment of bees with antibiotics is prohibited in the European Union (EU), significant progress being made in the EU risk assessment legislation (Barganska et al., 2011).

Starting with 2000, for the EU food and products intended for marketing, it became necessary to establish a maximum residue limit (M.R.L.), paving more attention to the negative effects produced by the residues or their metabolites that are found in products intended for human consumption. For this purpose, analytical methodologies were developed for identification and quantification of these compounds (Lazăr et al., 2006). Techniques for extraction and purification of antibiotics from animal origin samples (including honey) include some forms of liquid-liquid extraction (L.L.E.) or solid phase extraction (S.P.E.). The most used technique for drug extraction is liquid-liquid chromatography. H.P.L.C. method (High Perfomance Liquid Cromatography) is the most extensive chromatographic method used for the antibiotic's analysis (Burian, 2011; Vlaic et al., 2018).

The major classes of antibiotics present in beta-lactams, honev are: amphenicols, tetracyclines, macrolides, aminoglycosides and fluoroquinolones. Beta-lactams are antibiotics used to treat bacterial infections, altering bacterial cell wall biosynthesis; for example: penicillin, ampicillin, cloxacillin, amoxicillin (Sapna et al., 2010). Amphenicols blocks the enzyme peptidyl transferase on the 50S ribosome. The most used are: thiamphenicol, florfenicol, chloramphenicol. Chloramphenicol is an antimicrobial with a carcinogenic potential and an unaccepted substance to be used on animals intended for human consumption, including beekeeping (Orso et al., 2015). Tetracyclines are used for the bacterial diseases' treatment of the bee brood: for example: oxytetracycline, chlortetracycline, tetracycline. The action spectrum resembles to the one of chloramphenicol. Macrolides include about 40 antibiotics, among which the most known are erythromycin, tylosin, oleandomycin and spiramycin. There are two groups: macrolides with 14 carbon atoms

(erythromycin, oleandomycin) and macrolides with 16 carbon atoms (tylosin, spiramycin) (Mărghitas et al., 2010). The most known aminoglycosides are gentamicin, lincomycin, neomycin and streptomycin. The polar nature of these macrolides makes it difficult to isolate them from the samples and determine their chromatography (Barganska et al., 2011). Fluoroquinolones are used growth as promoters: example: for ciprofloxacin, enrofloxacin. norfloxacin. Organochlorine **pesticides** are highly toxic chemical substances used in agriculture to destroy pests. The presence of pesticide residues in honey has needed the establishment of monitoring programs to determine the human exposure. Many studies have shown that organochlorine pesticides accumulate in plants from polluted soil and can enter the food chain not only through fat products but also through non-fat poducts, such as honey (Panseri et al., 2014).

Organophosphorus compounds have been used as pesticides for almost five decades. They continue to be used as insecticides. helminthicides, ascaricides, nematocides and to a lesser extent as fungicides and herbicides. Although they have been and continue to be extremely useful in combating agricultural pests around the world, their widespread use had led to numerous poisonings, even with human victims. The primary acute toxicity to mammals associated with exposure to organophosphorus pesticides results from the acetylcholinesterase inhibition enzyme (Sultatos, 2009).

In Europe, other more commercial products are used by beekeepers to control varroosis: **amitraz**, coumaphos, fluvalinate and thymol (Faucon et al., 1995). *Varroa destructor* is a hematophagus ectoparasite of bees and it is considered to be a major cause of bee colonies loss in Europe and North America (Surlis et al., 2018).

MATERIALS AND METHODS

In this paper the aim was to evaluate the contamination degree of honey sourced from beekeepers from the centre and Southern Romania in order to form a large and homogeneous batch). The analyzes were performed in a laboratory, external of the processing unit. The study was conducted in the summer of 2019, on 3 batches of honey, collected by a local processor. The first batch consisted of 15 acacia honey samples from different beekeepers, analyzed before the honey homogenization process, the second one also consists of the same number and type of honey samples from other beekeepers from Romania, analyzed after the homogenization process, and the third lot consists of 15 polyfloral honey, each sample being harvasted from different beekeepers and .

The Nitroimidazole residues determination (metronidazole, dimetridazole, ronidazole) was carried out using the quantitative LC-(liquid chromatography-MS/MS method tandem spectrometry) (European mass regulation 37/2010/UE). Over the years, the LC-MS systems suffered significant changes, starting from simple analyses and reaching very accurate qualitative and quantitative analyses (Burian V., 2011). According to 470/2009/CE and 37/2010/CE regulations, the use of antibiotics in beekeeping is not allowed. The quantification limit of the method for metronidazole is 0.5 µg/kg, for dimetridazole is 2.5 μ g/kg and for ronidazole is 0.5 μ g/kg.

The tetracycline residue determination (oxytetracycline, tetracycline, doxycycline, chlortetracycline, demeclocycline, methacycline, minocycline) was conducted using the quantitative LC-MS/MS method. The quantification limit of the method is of 2 ppb, and according to the (EC) No 470/2009/CE and (EC) No 37/2010/UE Regulations, the use of antibiotics in beekeeping is not allowed.

residue The macrolide determination josamycin, (clindamycin, erythromycin, kitasamycin, lincomycin, oleandomycin, spiramvcin. mirosamycin, tilmicosin. trimethoprim, tylosin) was carried out using the LC-MS/MS method. For these, there is no legal limit, because the use of antibiotics in beekeeping is not allowed. The quantification limit of this method is 2 ppb (Regulation (EC)) No 37/2010/UE).

The Nitrofuran metabolites determination (semicarbazide from nitrofurazone, AOZ from furazolidone, AHD from nitrofurantoin) was conducted using the LC-MS/MS method. These substances are prohibited according to the Regulation (EC) No 37/2010/UE. The quantification limit of the method used is 1 µg/kg (Regulation (EC) No 2003/181/CE).

The chloramphenicol determination was conducted using the ELISA method. This is an officially appoved method. In accordance with Regulation (EC) No 2002/657/CE, up to 5% false negative results may occur. Chloramphenicol is a prohibited substance according to Regulation (EC) No 37/2010/UE. The quantification limit of the sample is 0.1 ppb (Regulation (EC) No 2003/181/CE).

The streptomycin and dihydrostreptomycin residues detection were carried out using the LC-MS/MS method. For these, there is no legal limit, because the use of antibiotics in beekeeping is not allowed. The quantification limit of this method is 2 ppb (Regulation (EC) No 37/2010/UE).

sulfonamides and trimethoprim The detection was made using the LC-MS/MS method. There were determined: sulfaquinoxaline, sulfadimethoxine. sulfamethizole. sulfachlorpvridazine, sulfamoxole, sulfadoxine, sulfasalazine, sulfabenzamide, sulfaguanidine, sulfanilamide, sulfacetamide. sulfadiazine. sulfathiazole. sulfapyridine, sulfamerazine, sulfamethazine, sulfamethoxypyridazine, sulfamethoxazole, trimethoprim, sulfamonomethoxine, sulfaclozine. sulfisoxazole. succinvlsulfathiazole, sulfaphenazole. sulfisozole, sulfisomidine. The quantification limit of the method is between 0.5-2 μ g/kg. For these, there is no legal limit, because the use of antibiotics in beekeeping is not allowed (Regulation (EC) No 37/2010/UE).

The Glyphosate residues determination was conducted using LC-MS/MS method. The quantification limit of this method is 0.010 mg/kg, the maximum residue level allowed is 0.050 mg/kg (Regulation (EC) No 369/2005/UE).

RESULTS AND DISCUSSIONS

Results and discussions regarding the contamination degree of the lot 1

The analysis of the 15 acacia honey samples, which form the first batch of the present study, the following results were obtained: **Results and discussion regarding the nitroimidazole residues determination:** following the method used, LC-MS/MS, for the 15 acacia honey samples, there were obtained values below the limit of quantification, respectively: metronidazole < 0.5 μ g/kg, dimetridazole < 2.5 μ g/kg, ronidazole < 0.5 μ g/kg. Considering the limit of quantification indicated above, this result was in accordance with Regulation (EC) No 37/2010/UE.

and discussion Results regarding the tetracvcline residues determinantion: regarding the results obtained from the analysis of the acacia honey samples by the LC-MS/MS method, the values obtained are below the limit of quantification, respectively $< 2 \mu g/kg$. Considering the limit, the result was in accordance Regulation (EC) No with 37/2010/UE.

Results and discussion regarding the macrolide residues determination: following the test of the 15 acacia honey samples by the LC-MS/MS method, the results are below the limit of quantification (< 2 μ g/kg). Therefore, the result was in accordance with Regulation (EC) No 37/2010/UE.

Results and discussion regarding the nitrofuran metabolites determination: the results obtained by LC-MS/MS testing of the 15 acacia honey samples are below the limit of quantification, respectively ($< 1 \mu g/kg$). The result was in accordance with the Regulation (EC) No 37/2010/UE.

Results and discussion regarding the chloramphenicol determination: following the analysis of the 15 acacia honey samples by the ELISA method, the result obtained is bellow the quantification limit of the method (< 0.1 μ g/kg). Considering the limit, the result was in accordance with Regulation (EC) No 37/2010/UE.

Results and discussion regarding the streptomycin and dihydrostreptomycin residues detection: the results obtained from the analysis of the acacia honey samples by the LC-MS/MS method regarding the detection of streptomycin and dihydrostreptomycin

residues, are below the limit of quantification (2 ppb). Considering the limit, the result was in accordance with Regulation (EC) No 37/2010/UE (regarding the residues of pharmacologically active substances in food products of animal origin).

Results and discussion regarding the sulfonamides and trimethoprim detection: regarding the analysis of the acacia honey samples by the LC-MS/MS method, the following average results were obtained (Table 1):

Table 1. Results and discussion regarding the sulfonamides and trimethoprim detection

sulfonamides and trimethoprim detection				
Analyzed parameter in µg/kg	LOQ*	Result		
Sulfadimethoxine	0.5	n.n. **		
Sulfaquinoxaline	0.5	n.n.		
Sulfamethizole	1	n.n.		
Sulfachlorpyridazine	2	n.n.		
Sulfamoxole	1	n.n.		
Sulfadoxine	0.5	n.n.		
Sulfasalazine	2	n.n.		
Sulfabenzamide	0.5	n.n.		
Sulfaguanidine	2	n.n.		
Sulfanilamide	2	n.n.		
Sulfacetamide	2	n.n.		
Sulfadiazine	1	n.n.		
Sulfathiazole	0.5	n.n.		
Sulfapyridine	1	n.n.		
Sulfamerazine	1	n.n.		
Sulfameter	1	n.n.		
Sulfamethazine	1	n.n.		
Sulfamethoxypyridazine	0.5	n.n.		
Sulfamethoxazole	1	n.n.		
Trimethoprim	0.5	n.n.		
Sulfamonomethoxine	0.5	n.n.		
Sulfaclozine	2	n.n.		
Sulfisoxazole	1	n.n.		
Succinylsulfathiazole	2	n.n.		
Sulfaphenazole	2	n.n.		
Sulfisozole	2	n.n.		
Sulfisomidine	1	n.n.		
*I $OO = limit of quantification:$				

*LOQ = limit of quantification;

**n.n. = below the limit of quantification.

Considering the limit, the result was in accordance with Regulation (EC) No 37/2010/UE.n

All the results obtained from the analysis of the batch1 (Table 2) werein accordance with Regulation (EC) No 37/2010/UE.

Performed analyses	Result	Allowed limit
β - γ -amylase activity	2.9 U/kg	Max. 5 U/kg
Nitroimidazoles	n.n.** (0.5-2.5 μg/kg)	MRL*
Tetracyclines	n.n. (2 μg/kg)	MRL
Macrolides	n.n. (2 μg/kg)	MRL
Nitrofuran metabolites	n.n. (1 μg/kg)	MRL
Chloramphenicol	n.n (0.1 μg/kg)	MRL
Streptomycin anddihydrostreptomy cin	n.n. (2 µg/kg)	MRL
Sulfonamides and Trimethoprim	n.n. (0.5-2 μg/kg)	MRL

Table 2. The obtained results regarding the contamination degree of batch 1 of acacia honey

*MRL = forbidden substance (Regulation (EC) No 37/2010/UE); **n.n. = below the limit of quantification.

Results and discussions regarding the contamination degree of the batch 2

After the analysis of the 15 acacia honey samples, which formed the batch 2, tested in the summer of 2019, the following results were obtained:

The results regarding the **glyphosate residues** determination are below the limit of quantification (0.010 mg/kg), the maximum allowed residue level being 0.050 mg/kg (Regulation (EC) No 369/2005/UE).

Regarding the results of the **nitroimidazole residues** determination, following the used method, LC-MS/MS, there were obtained values below the quantification limit in batch 2, respectively metronidazole < 0.5 μ g/kg, dimetridazole < 2.5 μ g/kg, ronidazole < 0.5 μ g/kg. Considering the limit of quantification, the result was in accordance with Regulation (EC) No 37/2010/UE (regarding the residues of pharmacologically active substances in food products of animal origin).

The **nitrofuran metabolites** determination, by the LC-MS/MS method showed values below the limit of quantification, respectively $(< 1 \ \mu g/kg)$.

Following the **sulfonamides and trimethoprim residues** detection, by the LC-MS/MS method, there were obtained results below the quantification limit of the method, respectively below 0.5-2 μ g/kg, depending on the analyzed parameter. Considering the limit of quantification, the result was in accordance with Regulation (EC) No 37/2010/UE (regarding the residues of pharmacologically active substances in food products of animal origin).

The chloramphenicol determination by the ELISA method, led to results below the quantification limit of the method (< 0.1µg/kg). Taking into account the quantification limit indicated previously, the result was in accordance with Regulation (EC) No 37/2010/UE (regarding the residues of pharmacologically active substances in food products of animal origin).

The results regarding the **streptomycin and dihydrostreptomycin residues** detection, by the LC-MS/MS method, are below the limit of quantification (2 ppb). Taking into account the quantification limit indicated previously, this result was in accordance with Regulation (EC) No 37/2010/UE.

All the results obtained from the analysis of the batch 2 (Table 3) werein accordance with Regulation (EC) No 37/2010/UE.

Table 3. The obtained results regarding the
contamination degree of batch 2 of acacia honey

0		5
Performed analyses	Result	Allowed
		limit
Glyphosate	n.n.**	0.050
	(0.010 mg/kg)	mg/kg
Nitroimidazoles	n.n.	MRL*
	(0.5-2.5µg/kg)	
Nitrofuran	n.n.	MRL
metabolites	(1 µg/kg)	
Chloramphenicol	n.n	MRL
-	(0.1 µg/kg)	
Streptomycin	n.n.	MRL
anddihydrostreptom	$(2\mu g/kg)$	
ycin		
Sulfonamides	n.n.	MRL
andtrimethoprim	(0.5-2 µg/kg)	

*MRL= forbidden substance (Regulation (EC) No 37/2010/UE); **n.n. = below the limit of quantification.

Results and discussions regarding the contamination degree of the batch 3

Following the analysis of the 15 polyfloral honey samples, which form the third batch of the present study, the following results were obtained:

Following the **glyphosate residues** analysis by the LC-MS/MS method, the obtained result is

below the limit of quantification (0.010 mg/kg), the maximum residue level allowed being 0.050 mg/kg (Regulation (EC) No 369/2005/UE).

The **chloramphenicol determination** by the ELISA method, led to values (maybe better in English) below the quantification limit of the method (< 0.1μ g/kg). Considering the limit of quantification, the result was in accordance with Regulation (EC) No 37/2010/UE.

Following the nitroimidazole residues determination (metronidazole, dimetridazole, ronidazole) for the 15 polyfloral honey samples, there were obtained values below the limit of quantification. respectively: metronidazole $< 0.5 \ \mu g/kg$, dimetridazole < 2.5 $\mu g/kg$, ronidazole < 0.5 $\mu g/kg$. Taking into account the quantification limit indicated previously, the result was in accordance with Regulation (EC) No 37/2010/UE.

The sulfonamides and trimethoprim residues detection by the LC-MS/MS method showed values below the quantification limit of the method, respectively below $0.5-2 \mu g/kg$, depending on the analyzed parameter. Considering the limit of quantification, the result was in accordance with Regulation (EC) No 37/2010/UE.

The **macrolide residues** are below the limit of quantification ($< 2 \mu g/kg$). Therefore, the result was in accordance with Regulation (EC) No 37/2010/UE.

For the **nitrofuran metabolites** determination (semicarbazide from nitrofurazone, AOZ from furazolidone, AHD from nitrofurantoin, AMOZ from furaltadon), there obtained results were below the limit of quantification, respectively ($< 1 \ \mu g/kg$), the result being in accordance with Regulation (EC) No 37/2010/UE.

Regarding the results of the **tetracycline residues** determinantion for the 15 polyfloral honey samples, the values obtained were below the quantification limit, respectively $< 5 \mu g/kg$. Considering the limit of quantification, the result was in accordance with Regulation (EC) No 37/2010/UE.

All the results obtained from the analysis of the batch3 were in accordance with Regulation (EC) No 37/2010/UE.

CONCLUSIONS

The results of toxic substances residues analysis (nitroimidazoles, tetracyclines, macrolides, nitrofuran metabolites, chloramphenicol, streptomycin, dihydrostreptomycin, sulfonamides and trimethoprim) for the acacia honey samples, which represent **Batch 1**, are below the limit of quantification.

Regarding Batch 2, consisting of 15 acacia honey samples, the value of glyphosate is <0.050 mg/kg and the residues of: nitroimidazoles. tetracvclines. macrolides. nitrofuran metabolites. chloramphenicol. dihydrostreptomycin, streptomycin. sulfonamides and trimethoprimare below the limit of quantification.

For the polyfloral honey samples which form the Batch 3, the residues of: nitroimidazoles, tetracyclines, macrolides, nitrofuran metabolites, chloramphenicol, streptomycin, dihydrostreptomycin, sulfonamides and trimethoprim are below the quantification limit. Following the conducted study on the 3 batches of honey, sourced from local beekeepers and analyzed in the summer of 2019, results were in accordance with the national and international legal requirements.

REFERENCES

- Barganska, Z., Namiesnik, J., Slebioda, M. (2011). Determination of antibiotic residues in honey. *TrAC Trend in Analytical Chemistry*, 30, 1035-1041.
- Burian (Bonta), V. (2011). Chromatographic techniques for determining contaminants in bee honey, University of Agronomic Sciences and Veterinary Medicine Cluj-Napoca, Doctoral School, Faculty of Animal Science and Biotechnology.
- Alquani, A.S., Owayss, A.A., Mahmoud, A.A. (2012). Mineral content and physical properties of local and imported honeys in Saudi Arabia, *Journal of Saudi Chemical Society*, 5, 618-625.
- Crivineanu, V, Goidea, A., Goran, G.V., Tudoreanu, L., Cretu, L.G. (2011). Findings on rheological and mineral parameters and heavy metal contamination of honeybees. *Lucrări Științifice Medicină Veterinară*, XLIV (1), 241-249.
- Dobre, I. R., Ghiţă, M., Fernoagă, C., Gajailă, G., Cotor, G. (2016). The principles and the regulation of organic beekeeping - conditions for ensuring the organic honey on the community market. *Journal of Biotechnology*, 231, S63.
- Dobre, I. R., Marmandiu, A., Gajaila, G., Cotor, G., Ghiță, M., Fernoagă, C. (2017). Prevention of

diseases in the beekeeping holdings–An essential condition for obtaining organic honey. *Journal of Biotechnology*, Volume 256, Page S68, ISSN 0168-1656.

- Doliș, M. (2009). *Beekeeping and sericulture / Apicultură și sericicultură*. Iași, RO: Alfa Publishing House.
- Faucon, J.P., Drajnudel, P., Fleche, C. (1995). Mise en evidence d'une diminution de l'efficacité de l'Apistanutilisécontre la varroase de l'abeille (*Apis mellifera* L). *Apidologie*, 26, 291-296.
- Lazăr Ștefan, Vornicu O.C., (2006). Beekeeping/Apicultura. Bucharest, RO: Alfa Publishing House.
- Mărghitaş, L.Al. (2010). Modern methods for determining residues and contaminants in honey and bee products/Metode moderne de determinare a reziduurilor şi contaminanților din miere şi produse apicole, Bucharest, RO: Academic Pres Publishing House.
- Murariu, F., Voda, A.D., Murariu, O.C. (2019), Researches on food safety assessment - supporting a healthy lifestyle for the population from NE of Romania, *Journal of Biotechnology*, 305, s68-s68.
- Murariu, O.C., Irimia, L.M., Robu, M., Işan, E. (2019), Ensuring nutrition security and sustainability of food systems as basis of human healhy life. *Proceeding of* the International Scientific Congress ",Life sciences, a challenge for the future", Filodiritto International Proceedings Editore, 175-180.
- Orso, D., Floriano, L., Ribeiro, L., Bandeira, L., Prestes, O., Zanella, R. (2015). Simultaneous Determination of Multiclass Pesticides and Antibiotics in Honey Samples Based on Ultra-HighPerformance Liquid Chromatography-Tandem Mass Spectrometry. *Food Analytical Methods*, 9, 1638-1653.
- Panseri, S., Catalano, A., Giorgia, A., Aricoli, F., Procopio, A., Chiesa, L. (2014). Occurrence of pesticide residues in Italian honey from different areas in relation to its potential contamination sources. *Food Control*, 38, 150-156.
- Petcu, C.D. (2006). *HACCP-Food safety guarantor*. Bucharest, RO: Idea Design Publishing House.
- Petcu, C.D. (2014a). Packaging used in the food industry/Ambalaje utilizate în industria alimentară. Bucharest, RO: Granada Publishing House.
- Petcu, C.D., Şulea, C., Dumitrache, M. (2014b). Audit of Producers/Users of Compressed Air and other Industrial Gases used in the Food Industry, *Quality-Access to Success*, 15 (130).
- Sapna, J., Nimisha, J. (2010). Antibiotics residues in Honey, New Delhi, Center for Science and Environment.
- Savu, C., Petcu, C.D. (2002). Hygiene and control of products of animal origin/Igiena şi controlul produselor de origine animală. Bucharest, RO: Semne Publishing House.
- Sultatos, L.G. (2009). Mammalian toxicology of organophosphorus pesticides. Journal of Toxicology and Environmental Health, 43, 271-289
- Surlis, C., James, C.C., Coffey, M., Kavanagh, K. (2018). Quantitative proteomics reveals divergent responses in *Apis mellifera* worker and drone pupae

to parasitization by Varroua destructor. Journal of Insect Physiology, 107, 291-301.

- Şapcaliu, A., Savu, V., Radoi, I., Tăpăloagă, D., Tanase, P., Calin, V. (2017). Evaluation of results in research made in order to obtain a phytotherapeutic product for the prophylaxis and fight against nosema in bees. *The EuroBiotech Journal*, 1(1), 36-40.
- Tamas-Krumpe, O., Bobiş, O., Mărgăoan, R., Chirilă, F., Lațiu, C., Ognean, L. (2019). Correlative research regarding the total polyphenolic content, antioxidant and antibacterial activity of three types of romanian honey. *Scientific Papers. Series D. Animal Science*, LXII (2), 282-288.
- Tofană, M. (2011). Food contaminants/Contaminanți alimentari, Cluj-Napoca, RO: Mega Publishing House.
- Tănăsoiu, I.C., Drăgotoiu, D., Drăgotoiu, T., Marin, M. (2014). Researches concerning the effects of supplementary feeding of bees families during autumn, winter, spring. *Scientific Papers. Series D. Animal Science*, LVII, 112-114.
- Tănăsoiu, I.C., Drăgotoiu, D., Marin, M., Drăgotoiu, T., Diniță, G. (2015). Research on the influence of eco certificated energo-protein use over the performance of bee families. *Agriculture and Agricultural Science Procedia*, 6, 265-271.
- Tăpăloagă, D., Tăpăloagă, P.R. (2017). Study regarding animal organic farming in Romania - current status and trends, *Scientific Papers. Series D. Animal Science*, LX.
- Tăpăloagă, D., Tăpăloagă, P.R. (2018), From conventional to organic agriculture - romanian past and future perspectives. *Scientific Papers. Series D. Animal Science*, 61(1), 239-244.
- Tudoreanu, L., Codreanu, M.D., Crivineanu, V., Goran, G.V. (2012). The quality of Romanian honey varieties - mineral content and textural properties. *Bulletin USAMV*, 69(1-2), 452-58.
- Vlaic, R.A., Muresan, A.E., Mureşan C.C., Petrut, G.S., Muresan, V., Muste, S. (2018). Quantitative analysis by HPLC and FT-MIR prediction of individual sugars from the plum fruit harvested during growth and fruit development. *Agronomy*, 8, 306, 1-16.
- Zacharis, K., Rotsis, I., Zachariadis, P.G., Zotos, A. (2012). Dispersive liquid-liquid microextraction for the determination of organochlorine pesticides residues in honey by gas chromatography-electron capture and ion trap mass spectrometric detection. *Food Chemistry*, 134, 1665-1672.
- Zacharis, K., Rotsis, I., Zachariadis, P.G., Zotos, A. (2012). Dispersive liquid–liquid microextraction for the determination of organochlorine pesticides residues in honey by gas chromatography-electron capture and ion trap mass spectrometric detection, *Food Chemistry*, 134, 1665-1672
- Commission Regulation (EC) 2002/657 laying down detailed rules for the application of Council Directive 96/23/EC on the operation of methods of analysis and interpretation of results.
- Commission Regulation (EC) 2003/181 laying down minimum performance limits (MRLs) for certain residues in food of animal origin.

- Regulation (EC) No 369/2005 of the European Parliament and of the Council on the maximum levels for pesticide residues in or on food and feed of plant and animal origin.
- Regulation (EC) No 470/2009 of the European Parliament and of the Council laying down Community procedures for the determination of

residue limits of pharmacologically active substances in foodstuffs of animal origin.

Commission Regulation (EU) 37/2010 on pharmacologically active substances and their classification according to maximum residue limits in foodstuffs of animal origin.