

THE BENEFICIAL EFFECT OF *BACILLUS* SPP. AS PROBIOTICS IN POULTRY NUTRITION - A REVIEW

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Abstract

Over the last few decades, the use of probiotics as source of feed additives in animal nutrition has increased considerably. As you know, sub-therapeutic levels of antibiotics were used as growth promoters (AGP) in the animal field, with extensive use in poultry industries, but due to their multivarious side effects, it was necessary to find some alternatives in order to satisfy the consumer's demands. Probiotics are considered one of the options as a significant alternative to antibiotics for improving health, growth, and poultry production. In our day, among the extensive number of probiotic products in use are bacterial spore formers, mostly of the genus Bacillus. The current review presents the benefits of probiotic utilization based on Bacillus spp. in poultry feed highlighting their potential to form spores that can withstand harsh environmental stress and transition during poultry gastrointestinal tract. Furthermore, Bacillus spores involve more than 80% survivability during the probiotic in vitro tests, remaining stable in a fairly high concentration. Based on the information found from published articles, this review summarizes stronger information about the properties of Bacillus spp. obtained from in vitro and in vivo screening, which can provide researchers with a better understanding of the use of this species in poultry nutrition.

Key words: *Bacillus spp., poultry, probiotics.*

INTRODUCTION

The use of antibiotics as growth promoters (AGP) in animal diets has been of concern and has even been banned in many countries globally, due to the appearance of resistant bacteria to antibiotics, which was associated with human and animal illnesses (Cartman et al., 2004; Bajagai et al., 2017; Jiang et al., 2021). Excessive antibiotic utilization involves the appearance and transfer of gene resistance between bacteria, the disequilibrium of normal microflora, and the decline of beneficial intestinal bacteria (Sinol et al., 2012). Moreover, Cervantes (2015) affirmed that the elimination of antibiotics involves considerable consequences with negative effects on performance production, infections of the gut, and the possibility of a high mortality rate in the poultry industry.

Since 2006, many European Union (EU) countries prohibited using all commonly feed antibiotics added as growth promoters (EC Regulation No. 1831/2003). The EU has included this issue as the main point of *“the*

farm to fork concept” and, shortly, the European Commission will act to decrease the total antimicrobials sales for livestock animals (European Commission, 2020). Therefore, probiotics are increasingly popular as considerable safe alternatives to replace and reduce antibiotics (Meng et al., 2010; Nguyen et al., 2018; Luise et al., 2022), as a viable solution to save the animal livestock sector (Hmani et al., 2017; Park et al., 2015), especially for young animals like broilers and piglets (Aar et al., 2016; Idriceanu et al., 2020). According to the Food and Agriculture Organization (FAO, 2002) and World Health Organization (WHO, 2006) guidelines, probiotic bacteria are an important solution and have been proved the most favoured alternative to antibiotics as AGP and inhibitor of pathogens in the animal industry (Zhang & Kim, 2014).

Probiotics are defined as a preparation containing viable or inactivated known bacteria (Ramlucken et al., 2020a) and generally are recognized as safe (GRAS). Also, the use of probiotics or direct in-fed microbial (DFM)

which, when administered in adequate amounts, confer “a health benefit to the host” (Fuller, 1989; Schrezenmeir & De Vrese, 2001) and seems to be one of the most promising strategies (Barba-Vidal et al., 2019).

Probiotics occur an important place due to their beneficial impact on body weight host, growth performance, improving the health profile (Abd El-Hack et al., 2020; Zhang & Kim, 2014), gut immunity by regulating the metabolism, and bacteria compositions from this area (Luise et al., 2022).

Generally, a probiotic strain is recommended to be isolated from the same source for which it was created. Based on the probiotic perspective, it is proclaimed that the candidate probiotic should be isolated from the source of the target population, which helps them to grow well inside the selected host (Elshaghabe et al., 2017).

Before a probiotic product can be included in poultry feed, it is essential to assay its stability (viability and growth) under simulation of the gastrointestinal tract (GIT) harsh conditions. As a level of inclusion, a probiotic must retain less than 1×10^6 CFU g^{-1} (Millette et al., 2013).

Probiotics improve digestion and nutrients absorption by inhibiting potentially pathogenic bacteria, regulating intestinal affection (Ding et al., 2021), and modulating the gut microbiota, which plays a critical role in sustaining beneficial health status (Patil et al., 2015). Also, probiotics addition as feed additives or supplements can re-establish the ecologic stability of gut microbiota by inhibiting pathogens and promoting the growth of representative bacteria (Bermúdez-Humarán et al., 2019; Del Toro-Barbosa et al., 2020).

An ideal probiotic is necessary to have the capacity to adhere to the intestinal mucosa, grow rapidly, and maintain its viability (Luise et al., 2022). Manufacturing is an additional trait of probiotics including transport and storage conditions, applied usually in the processes for obtaining animal feed, to keep as much possible the vital properties of these products (Banjagai et al., 2016), especially after feed pelleting, storage, and manipulation (Cutting, 2011).

The most commonly used probiotics are Gram-positive bacteria from the genus *Bacillus*, *Lactobacillus*, *Lactococcus*, *Streptococcus*,

Bifidobacterium, *Enterococcus*, and non-bacteria (yeast or fungal) including *Aspergillus oryzae*, *Candida pintolopesii*, *Saccharomyces boulardii*, and *Saccharomyces cerevisiae*, which are widely used to prevent poultry diseases, pathogens multiplication and improve the growth performance (Mountzouris et al., 2007; Gaggia et al., 2010; Elshaghabe et al., 2017; Kerry et al., 2018; Dumitru et al., 2020; Yoha et al., 2021).

Currently, through the large number of probiotic products, bacterial spore formers are in use today (Hong et al., 2005) and have been most extensively studied. Species from the *Bacillus* genus present a distinct advantage over other probiotics due to the capacity of sporulation (Kim et al., 2019), germination, and proliferation within the GIT of animals (Dumitru et al., 2019; Ciurescu et al., 2020; Dumitru et al., 2021).

As Gram-positive or Gram-variable rods, catalase producing and efficient probiotic product, *Bacillus* spp. is necessary to survive during environmental stress, preparation conditions and application processes, tolerance to low pH (Lee et al., 2017; Penalzoza-Vazquez et al., 2017), bile salts concentrations, and other severe conditions for the keep of their viability and properties within GIT (Barbosa et al., 2005; Shivaramaiah et al., 2011; Jiang et al., 2021; Dumitru et al., 2020). Morphologically, *Bacillus* species have rod-shaped cells with squared or rounded ends between 0.5×1.2 to $2.5 \times 10 \mu m$, occurring singly or in chains, and chains stability determines the colony form, which may differ from strain to strain (Logan & De Vos, 2009).

In comparison with other probiotic bacteria *Bacillus* spp. have notable advantages due to the capacity that are endospore-forming aerobic or facultative anaerobic bacteria. Sosa et al (2016) affirmed that *Bacillus* spp. under stressful environmental conditions can produce spores that remain in a dormant state for long periods (more than 2 years).

This trait makes them thermostable for storage and processing (i.e., extrusion and pelleting), with resistance to extreme temperatures up to $113^{\circ}C$ for 8 min (Grant et al., 2018). This property makes it easier to control and enhances its probability of surviving during the animal feed production process. Further,

Cartman et al. (2008) affirmed that *Bacillus* spp. can utilize nitrate or nitrite to facilitate anaerobic respiration, which enables them to survive in anoxic conditions.

Additionally, *Bacillus* spores were confirmed to survive at low pH in the stomach, bile salts, harsh conditions in the GIT environment of the host (Barbosa et al., 2005; Chaiyawan et al., 2010; Wang et al., 2010; Cutting, 2011; Bajagai et al., 2016; Dumitru et al., 2018; Dumitru et al., 2020), high pressures, and caustic chemicals, making them suitable for distribution and commercialization (Cartman et al., 2007).

Regarding the *Bacillus* group, the bacilli are easy to produce by conventional fermentation and do not involve expensive manufacture to ensure a stable commercial product (Cutting, 2011; Ramlucken et al., 2020b).

The addition of viable probiotics such as DFM, including bacteria from *Bacillus* group, involves beneficial health. Usually, *Bacillus* are examined as probiotic products in monogastric animals: *Bacillus subtilis*, *Bacillus licheniformis*, *Bacillus amyloliquefaciens*, *Bacillus coagulans*, *Bacillus cereus* and *Bacillus megaterium* (Cutting, 2011; Ciurescu et al., 2020; Dumitru et al., 2021; Uraisha et al., 2021; Mushtaq et al., 2022).

The anti-nutritional factor from feed materials raw could potentially be neutralized by using enzymes that occur a vital role in nutrient absorption by diminishing intestinal viscosity through catalyzing undigested starch polysaccharides (Popov et al., 2021).

Bacillus species possess several capacities such as secretion of beneficial enzymes (amylase, protease, cellulase, lipase, xylanase, phytase, and keratinase), antimicrobial molecules production (Ramlucken et al., 2020c; Sumi et al., 2015), and beneficial metabolites through modification of gut microflora (Grant et al., 2018; Dumitru et al., 2019; Shah & Bhatt, 2011; Jani et al., 2012). Further, the capacity of sporulation extends the percentage of survivability (heat tolerance, low pH of the gastric barrier, and longer viability during storage) in several environmental conditions compared to those containing non-spore-forming bacteria (Mingmongkolchai & Panbangred, 2018; Dumitru et al., 2021).

Bacillus improves the intestinal immune system by raising the levels of cytokines and chemokines as interleukin-1 β (IL-1 β) and interferon- γ (IFN γ) in the chicken gut (Lee et al., 2013).

In the last decade, the use of probiotics in animal feed has occupied significant attention, and the majority of the recent probiotics are represented by lactic acid bacteria, especially *Lactobacillus* spp. and *Enterococcus* spp. (Sorescu et al., 2019).

Based on published studies, this review will focus on the dietary supplementation with *Bacillus*-based probiotic in broiler chickens and the positive traits of this genus that has functional effects on the development of suitable commercial probiotics in poultry nutrition.

MATERIALS AND METHODS

To conduct this review, more than 160 references were necessary based on *in vitro* probiotic properties which establish the desirable *Bacillus* characteristics of several strains for survivability during GIT harvest conditions, and their effect as a probiotic product in poultry nutrition. The electronic search was carried out over the last 25 years from articles published in ISI Journals, Web of Science (WoS), and Scopus. For example, the systematic topic of research data was carried out from Google Scholar (<https://scholar.google.com>), ScienceDirect (<https://www.sciencedirect.com>), PubMed (<https://pubmed.ncbi.nlm.nih.gov/>), and NCBI-PCM (<https://www.ncbi.nlm.nih.gov/pmc/>). The topic of interest as a strategy for the search was based on probiotics effect on poultry nutrition. In addition, the keywords used for the search were: probiotics, alternative to antibiotics, probiotic properties, pH resistance, bile salts tolerance, immune response, spores viability, enzymatic activity, microflora, intestinal health, broiler performance, *Bacillus* spp., and poultry nutrition.

RESULTS AND DISCUSSIONS

Probiotics history, niche and mode of action

Elie Metchnikoff was the first investigator in the fermentation processes field and probiotic products. He reported that large soured milk

consumption increases human longevity. Furthermore, Metchnikoff affirmed that the lower gut can be affected by microbes, generally bacteria from *Lactobacillus* genus, for instance, *L. bulgaricus* (Ran et al., 2019). The term probiotic is correlated with “life” being considered “microbial feed supplements that can affect positively the host”. Over the years, the meaning of probiotics was changed. Later, in 1953, Werner Kollath gave other terminology as, “probiotika” and defined as “live microorganisms which are essential for the healthy development of the gut for life”. The definition of probiotic was in continuous modification. Lilley and Stillwell (1965) defined probiotics as possible microorganisms with the capacity to help the proliferation of another beneficial microorganism. Our days, their definition is opposite to the antibiotic terms (Abd El-Hack et al., 2020). Morelli & Capurso (2012) defined probiotics as the consumption of enough live microorganisms with the capability to contribute health benefits

to the host. Also, the authors affirmed that some strains ingested by the host may induce other reactions in the body. An example that can be given is related by *Bifidobacterium* spp. which can produce metabolic end products (acetate and lactate) with the capacity to diminish Gram-positive and Gram-negative pathogenic bacteria (Abd El-Hack et al., 2020). *Bacillus* species have been isolated from a diversity of habitats as soil, vegetables, water, animals, and as a transient part of the human gut, contaminants of raw and prepared foods, aviation fuels (Kotb, 2014; Alou et al., 2015), feces from different animals as chickens, pigs, ruminants and aquatic animals (Mingmongkolchai & Panbangred, 2018). Hong et al. (2005) affirmed that species from *Bacillus* are normally allochthonous from GIT due to the ingestion of bacteria from soil and contaminated food. Table 1 is presented *Bacillus* probiotics isolated from different sources and their benefits in the poultry industry.

Table 1. *Bacillus* isolation from diverse sources with applicability in poultry

Bacillus designation	Sources	Benefits	Reference
<i>B. subtilis</i>	Soil	Improve the growth performance, gut, excreta bacterial community, immune system and gut health, regulate intestinal microstructure and digestive enzymes.	Bar & Friedman (2018); Ciurescu et al. (2020); Liu et al. (2020); Oladokun et al. (2021).
<i>B. subtilis</i> CH16	Chicken GIT	Increase in daily weight gain (ADG), body weight (BW) and biofilm formation, reduce feed conversion ratio (FCR).	Nguyen et al. (2015)
<i>B. subtilis</i> (SC2362, 1781, 747, ATCC PTA-673, PB6)	Environment sources, soil	Capacity to germinate in GIT. Increase eggshell thickness, decrease excreta <i>Salmonella</i> counts without harmful effect on performance. Beneficial influence on selected performance parameters, egg quality, and the cholesterol content of yolk lipids. Greater resistance to the avian pathogenic <i>Escherichia coli</i> O78:K80 with a reduction in the colonization of the spleen, liver, and caeca.	Cartman et al. (2008); Sobczak & Kozłowski (2015); Park et al. (2020); La Ragione et al. (2001); Jayaraman et al. (2017)
<i>B. subtilis</i> fmbJ	Soil	Significantly decreased reactive oxygen species (ROS) contents in liver mitochondria of broilers.	Bai et al. (2017)
<i>B. subtilis</i>	Soil	Release antimicrobial and antibiotic compounds.	Jayaraman et al. (2017)
<i>B. subtilis</i> 1781 (PB1): <i>B. subtilis</i> 1104+ <i>B. subtilis</i> 747	Environment sources	Modify intestinal activity and influence gut barrier integrity through increased tight junction gene expression.	Gadde et al., (2017b)
<i>B. licheniformis</i>	Unknown	Enhance meat, necrotic enteritis, and enhance growth performance.	Liu et al. (2012); Cheng et al. (2017)
<i>B. subtilis</i>	Soil	Improve feed conversion efficiency and diminish abdominal fat. Reduce the intestinal size and promote the growth of several digestive organs.	Samanya et al. (2002); Wang et al. (2018)
<i>B. amyloliquefaciens</i>	Soil	Increase serum immunoglobulin levels, decrease the number of <i>E. coli</i> , NH ₃ , and H ₂ S emissions. Enhances gut health and growth performance.	Ahmet et al. (2014); Tang et al. (2017); Li et al. (2015)

<i>B. coagulans</i>	Soil	Protective efficacy in <i>Salmonella enteritidis</i> infections.	Zhen et al. (2018)
Symbiotic: prebiotic (xylooligosaccharide and yeast) and probiotic (<i>B. licheniformis</i> , <i>B. subtilis</i> and <i>C. butyricum</i>)	Soil	Prevents necrotic enteritis and enhances growth performance.	Li et al. (2019)
<i>B. subtilis</i> KATMIRA1933 and <i>B. amyloliquefaciens</i> B-1895	Soil	Positively affected egg production, quality of sperm, quality and hatchery of eggs.	Mazanko et al. (2018)
BioPlus 2B (preparation of <i>B. subtilis</i> DSM 5749 and <i>B. licheniformis</i> DSM 5750)	Soybean mash and soils	Improve growth performance.	EFSA (2019)
<i>B. subtilis</i> and <i>B. licheniformis</i>		Performance improvement and control effects of <i>Salmonella</i> infection.	Abudabos et al. (2020)
<i>B. pumilus</i> and <i>B. subtilis</i>		Beneficial effects the intestinal and immune activities, specifically in day-14.	Bilal et al. (2021)

The presence of spores makes the *Bacillus* group to resist in extreme conditions (stomach acidity, bile salts concentrations etc.). Besides, during processing and storage, the bacilli spore-formers involve more stability, making them suitable as an ingredient for probiotic formulations (Elshaghabee et al., 2017). Different supposable mechanisms for probiotic action have been investigated based on inhibition and stimulation of the host immunity (Guo et al., 2020). The main interest in animal nutrition is occurred to the relationship between

nutrition and gut health, mainly in the small intestine (Luise et al., 2022). An important characteristic of this genus is its rapidity of growth and capacity for survival during chickens' GIT (Lattore et al., 2014). The mechanisms of *Bacillus* spp. action in their vegetative state may function as probiotics are similar to those of other probiotic organisms (Ramlucken et al., 2020c). Figure 1 is illustrated the main actions of *Bacillus* strains in the organism host.

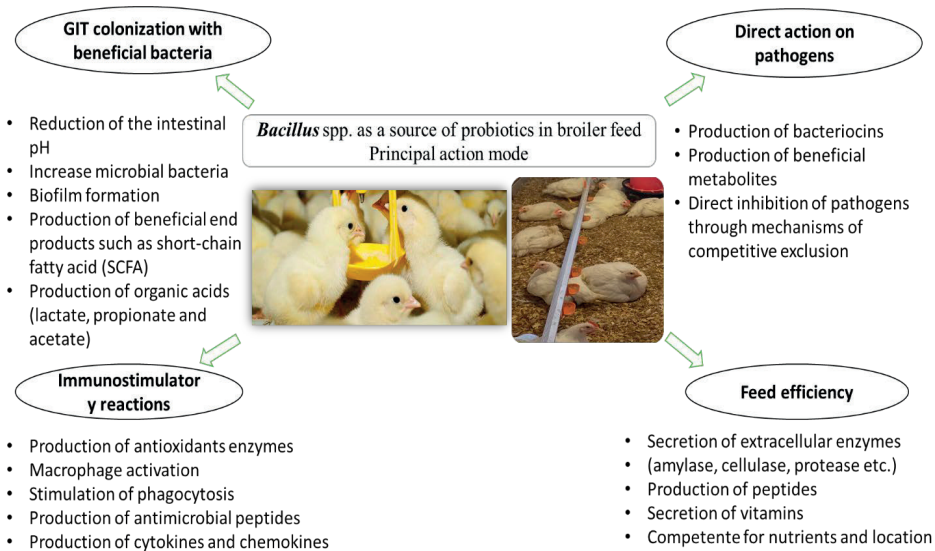


Figure 1. The effects of *Bacillus* as a probiotic source in poultry nutrition

It is known that the oral cavities of birds do not present teeth, compared to the mammals, so, feed going down to the esophagus into the crop (Scanes, 2014). Saliva secretion starts the process of feed humidification in the oral cavity involving a moist and good medium for

the progress of bacteria in the crop (Scanes, 2014).

As a group of bacteria *Lactobacillus*, *Enterobacterium* and *Bifidobacterium* were most representative in the broiler chicken crop (Feye et al., 2020). Additionally, after a review

by Feye et al. (2020) large number of *Bacillus* (more than 70%) can be found in the poultry crop. In this region of the GIT, the feed can remain around 14 h, but many times stay between 1-3 h (Scanes, 2014).

It is very important when using a probiotic product based on *Bacillus* to be resistant to harvest environment conditions. Due to the low pH of the glandular stomach (2.3-4.8) of chickens, the probiotic of interest should not resist (Grist, 2006) which is why most of the microorganisms numbers are lower compared to the crop and intestine. Furthermore, *Bacillus* species have the capacity to produce extracellular enzymes; the first part of the enzymatic process of digestion of feed begins in the gizzard, the organ where feed is broken down and transported to the small intestine in small portions (Scanes, 2014).

The small intestines of poultry (duodenum, jejunum, and ileum) present a gradual pH from 5.0-6.0 (Ciurescu et al., 2020); however, in the small intestine, the feed remains 2 to 8 h, a process that is accompanied by secretion of enzymes and mucin (Scanes, 2014), digestion and absorption of nutrients.

A study by Dumitru et al. (2019) demonstrated the *B. licheniformis* capacity to produce extracellular enzymes by determining the carbohydrate fermentation profile through API 50CHB kits, resistance at low pH (2.0 and 3.0), and bile salts concentrations, properties that are necessary to be evaluated before administration of DFM probiotics in animal nutrition.

In the cecum, feed stays for 12-20 h (Oakley et al., 2014; Clavijo & Flórez, 2018). Gang et al. (2002) affirmed that the first function as a result of fermentation in the cecum is enzymatic activity and detoxification of damaging substances.

As mentioned above, the conditions during GIT are different, and not every bacteria can resist. In this case, the best option is to find significant probiotics through the commensal microorganisms that populate the interest intestine area (Popov et al., 2021).

Competition for adhesion to the intestinal epithelium

The selection of *Bacillus* as DFM candidate or probiotic product is based on adhesion to the epithelial surface, colonization, and population

of the GIT host, and afterward to form a strong barrier to prevent the adhesion of pathogens (Chauhan & Singh, 2019).

The results obtained by Nishiyama et al. (2020) suggest that *B. subtilis* C-3102 supplementation presented the potential to diminish *S. enterica* infection rates and accelerate the pathogen exclusion from the cecum, spleen, and chickens liver. Another study confirmed the potential of *B. amyloliquefaciens* US573 for exhibiting good adhesion efficacy to chicken enterocytes and the ability to create biofilms that may favour survivability in the animal tract. Moreover, the US573 strain neutralizes the antinutritional factor and maximizes nutrient absorption due to the enzymatic activity (xylanase, β -glucanase, and amylase). Therefore, the mode of action of poultry probiotics is not very clear, for this reason, further studies with *Bacillus* as DFM-probiotics will be in continuous research.

Secretion of inhibitory substances

Bacillus species are well-known producers of antimicrobial peptides such as bacteriocins, small ribosomal peptides with the capacity to inhibit the growth of pathogens. Khalique et al. (2020) showed that *B. subtilis* SP6 exhibits a wide antibacterial spectrum that has antagonist activity against *Clostridium perfringens*, a normal inhabitant in chicken intestinal microflora, usually found in low numbers in the posterior gut section (Arif et al., 2021).

Bacteriocins can attend as colonizing peptides by facilitating the population of probiotic strains into an already employed niche on the intestinal epithelium (Bahaddad et al., 2022). *B. subtilis* KATMIRA1933 and *B. amyloliquefaciens* B-1895 are probiotic strains that secrete the bacteriocins subtilisin A and subtilin which have significant potential against *Salmonella* by inhibition of biofilm formation which may serve in decreased the pathogens microorganisms (Tazehabadi et al., 2021).

Modulation of the immune system

It is known that innate immunity is the first line of protection against pathogens. The probiotic spore-forming bacteria have been described for their aptitude to stimulate and/or control the

poultry immune system by secreting cytokines and immune defence substances (Popov et al., 2021).

Probiotic supplementation, as an immunomodulatory answer, positively increased the level of serum immunoglobulin (Paturi et al., 20007). In addition, Mountzouris et al. (2007) affirmed that the inclusion of probiotics stimulates GIT immunity by decreasing the number of pathogens in microflora. Likewise, Xu et al. (2012) reported that *B. subtilis* stimulated the production of cytokines as Il-10 and Il-4. Furthermore, *Bacillus* spp. registered a vital role in cytokines regulation (Mushtaq et al., 2022), immune modulation, and activation of macrophages without cytotoxicity (Popov et al., 2021).

According to Bai et al. (2017), the inclusion of *Bacillus* spp. in broiler diets improves radically the IgA. Al-Khalaifa et al. (2019) reported that the administration of probiotics in broiler production improves significantly the immune globulin. Furthermore, Fathi et al. (2017) described those dietary probiotics in broiler chickens improved the level of immunoglobulins (IgA, IgM, and IgG).

Several studies were demonstrated that the inclusion of *B. subtilis* in chickens' diets enhances the innate and acquired immune responses of broiler (Pagnini et al., 2010; Lee et al., 2015; Gadde et al., 2017a; Guo et al., 2020; Tarradas et al., 2020; Sikandar et al., 2020). Besides, *B. subtilis* was shown to modulate the responses of immune protective hosts against potential infections (Rajput et al., 2014; Zhang et al., 2017).

Spore formers as probiotic product

Due to physiological properties, *Bacillus* species have the capacity to produce a multitude of enzymes, metabolites, antibiotics, having thus a high spectrum of utilization in medical and pharmaceutical fields, agricultural and industrial processes, animal nutrition, etc. (Celandroni et al., 2019).

The most common bacteria from the *Bacillus* group used as a probiotic product in animal production including in the poultry industry is the *B. subtilis* strain (Joerger & Ganguly, 2017; Idriceanu et al., 2020).

Efficacy of probiotic inclusion can be ascribed to the species of bacteria and the formula of

supplementation used, such as wet (liquid culture) or powdered (lyophilization) (FAO & WHO, 2001). Administration of *Bacillus* as a probiotic product in the poultry diet can be performed orally, directly in water drinker (as liquid inoculum culture), or homogenized along with the feed (Lei et al., 2015; Lattore et al., 2017; Ma et al., 2018; Ciurescu et al., 2021). During the manufacturing including fermentation, drying, freezing, thawing, and rehydration, *Bacillus* spores have the ability to resist passage through the GIT, proliferate and populate the host digestive tract (Elisashvili et al., 2019; Popov et al., 2021).

Bacillus is a group recognized as spore-forming bacteria, known for their capability to germinate, proliferate, and re-sporulate. Due to the production of endospores, Bacilli involve long viability making them more stable and resistant to harvest environmental conditions. Keller et al. (2020) showed that when used *in vitro* human model, the *B. coagulans* GBI-30 can proliferate up to 97% in the GIT with active metabolically cells. Lattore et al. (2014), also, noted that 90% of *Bacillus* spores germinate in the small intestine of chickens within one hour. The spores can adhere to the intestinal walls, germinate and sporulate under anaerobic conditions as commensal to the animal intestines (Hong et al., 2009; Auger et al., 2009).

Probiotic benefits in poultry diets

The administration of probiotics in the poultry field specifically in broiler chickens has positive impacts on growth performance, feed efficiency, gut histomorphology improvement, immunity status, increase diseases resistance, and a beneficial microbiota increment (Simon et al., 2001; Mountzouris et al., 2010; Grant et al., 2018; Zhen et al., 2018).

As a direct effect, *Bacillus* probiotics can act on pathogenic bacteria such as *E. coli*, *Salmonella*, *Clostridium*, and *Campylobacter*, ensuing inhibition of their growth and population of the animal gut (Luise et al., 2022), thus preventing or reducing the incidence of infections (Zhang et al., 2014; Ding et al., 2017; Castaneda et al., 2021). For example, necrotic enteritis (NE) in the broiler industry, is produced by *Clostridium perfringens*, which is a digestive tract infection

with negative effects on host profitability (Abd El-Hack et al., 2022) and has conducted over the years to economic losses (Salem and Attia, 2021). As an enteric disorder, *C. perfringens* can be found in air, wastewater, healthy human, and animal GIT (Khelifa et al., 2012). Through 2 and 6 weeks, NE can occur in the broiler chickens tract due to the strong characteristics caused by *C. perfringens* pathogen (anaerobic, Gram-positive, endospore-forming, without motility), which could survive and stay life in extreme environmental conditions such as disintegration of organic matter and soil, due to the capacity to form endospores (Khelifa et al., 2015).

Studies have presented that inclusion of *Bacillus* spp. improves overall intestinal health and performance growth in broiler chickens (Grant et al., 2018). Teo & Tan (2007) showed that two types of *B. subtilis* strains, isolated from the chicken gut, involved antagonistic action against *C. perfringens* ATCC 13124. Later, in 2010, Knap et al. observed a reduction of *C. perfringens* in chickens at the addition of three levels of *B. licheniformis* s (8×10^5 CFU/g feed, 8×10^6 CFU/g feed, and 8×10^7 CFU/g feed). All three concentrations maintained similar body weight (BW) and feed conversion ratio (FCR) in the chicken's trial. Further, intestinal *Salmonella typhimurium* was significantly decreased in the presence of *B. subtilis* B2A (1×10^4 CFU/g, 1×10^5 CFU/g, and 1×10^6 CFU/g). Park and Kim (2014) reported a better feed conversion ratio (FCR) and a less feed intake (FI) in the experimental chicken group feed with *B. subtilis* B2A. As mentioned, *Bacillus* is in a vegetative state it is possible to not persist in the chicken's intestinal epithelium for a long time (Latorre et al., 2014). However, *Bacillus* once inside, the chicken's GIT germinates rapidly and vegetative cells can outnumber spores within 20 h of oral administration as mentioned by Cartman et al. (2008). The presence of spores could be detected over GIT. So, in the gut wall, bacteria from *Bacillus* group start to colonize these host section which competes with and block the pathogenic bacterial sites (Mushtaq et al., 2022).

The pathogens prevention could be due to the secretion of antimicrobial peptides by *Bacillus* spp. such as amylase and protease enzymes

(Dumitru et al., 2018) and metabolites (lipopeptides, surfactins, bacteriocins, inhibitory substances) which involve antagonistic results for microorganisms (Baruzzi et al., 2015; Sumi et al., 2015). It is known that, when an enzymatic bacterium is added to animal feed, the absorption and nutrient availability will improve (Amerah et al., 2017).

During competition with pathogens from the gut, the host can recover a part of the energy lost by captivating nutrients and metabolites such as lactic acid and volatile fatty acids resulting from fermenting bacteria (Grant et al., 2018).

Utilization of nutrients

An effective probiotic for growth and proliferation within the host is necessary to use nutrients and energy (Jha et al., 2020). The effects of the inclusion level of five probiotic bacterial strains (*L. reuteri* DSM 16350, *L. salivarius* DSM 16351, *Enterococcus faecium* DSM 16211, *Bifidobacterium animalis* DSM 16284, and *Pediococcus acidilactici* DSM 16210) were investigated by Mountzouris et al. (2010) as sources of probiotics in broilers feed (hybrid Cobb, male). The author's study found that the higher inclusion level ($> 10^9$ CFU/kg feed) modulates the cecal microflora composition and enhanced the growth performance and nutrient utilization in a corn-soybean diet.

Higher digestive absorption of nutrients in animal nutrition in the presence of probiotic supplementation is followed by an improvement of the intestinal structure and environment gut modulation (Choi et al., 2011). For example, the place of proliferation and differentiation of intestinal epithelial cells that stimulate villus growth is occurred by crypts.

The addition of *B. subtilis* DSM 29784 improved interior eggs quality with a significant increase in nutrient retentions and dietary apparent metabolizable energy (AME) in laying hens during the production cycle (Neijat et al., 2019).

Among of four inclusion levels (100, 150, 200, and 250 mg/kg, time 42 days) of *B. subtilis* improved the apparent metabolism of crude protein, crude fat, dry matter, and organic matter. Overall, Gao et al. (2017)

recommended that the inclusion of *B. subtilis* at 200 mg/kg could improve the broiler's performance.

He et al. (2019) investigated the effect of *B. subtilis*, *B. licheniformis*, and *S. cerevisiae* as an antibiotic substitute on growth performance and intestinal health status in broilers. The results obtained showed that the inclusion of probiotic complex as an alternative to chlortetracycline could improve performance growth, nutrient digestibility, serum antioxidant capacity, jejunal mucosal barrier function, and intestinal broilers morphology.

Even in unsuitable farming conditions, such as spore-forming bacteria, *B. subtilis* is implied in improving nutrient digestibility (Jha et al., 2020). After intramuscular inoculation with *E. coli*, the control + 0.1% *B. subtilis* increased the digestibility of nutrients ($P < 0.01$) followed by a reduction of *E. coli* respectively broiler colibacillosis disease (Manafi et al., 2017).

A large number of microorganisms are used as probiotics in poultry; the inclusion of *B. subtilis* DSM 17299 in broilers was correlated well with high nutrient digestibility of dry matter (DM), crude protein (CP), and AME (Reis et al., 2017). Due to their capacity to synthesize enzymes, *Bacillus* species occur as an essential solution in animal nutrition. Several studies reported that dietary probiotic supplementation enhanced the ileal digestibility of some nutrients like CP and most amino acids (Apatha, 2008; Oso et al., 2019).

Due to their intense activities, digestive enzymes can affect nutrient digestibility (Zaghari et al., 2016). The improvement of digestibility, nutrients absorption, and digesta viscosity may be associated with the production of extracellular enzymes by the vegetative form of *B. subtilis* which secrete protease, amylase, and lipase (Chen et al., 2009) improving, in the end, the productive animal profile (Ravindran, 2013). The addition of *B. subtilis* enhances the feed efficiency, highlighting the production level through oxygen utilization in host GIT and secretion of various enzymes like subtilisin and catalase (Bajagai et al., 2016).

Guo et al. (2020) proved that *B. subtilis* used in their study had the capacity to produce protease. Another study affirmed that *B. licheniformis* ATCC 21424 was found as an effective enzyme producer (amylase and

protease) through the submerged fermentation process (Dumitru & Habeanu, 2021). Enzymes supplementation aims to reduce the presence of indigestible components and can subsidize better digestion and therefore intensify the nutritional value of feed and energy in animal nutrition (Hmani et al., 2017).

Health status

According to the literature data, the administration of *Bacillus* diminishes the broiler chickens mortality (Teo and Tan, 2007; Knap et al., 2010; Abdel Baset et al., 2020; Qiu et al., 2021). In addition, *B. subtilis* used as probiotic (8×10^5 CFU/g) in poultry feed decreased mortality by 2.51% compared with non-supplemented groups (Harrington et al., 2016). Indeed, due to the possibility to germinate, with rapid multiplication, *Bacillus* strains have more advantages providing a wide range of health benefits to the host. Sen et al. (2010) reported that the addition of *B. subtilis* LS 1-2 improved the intestinal microbial balance and gut health of broiler with a decrease in cecal *Clostridium* and Coliforms counts. Further, *B. amyloliquefaciens*-based DFM as replace of antibiotics decreased the cecal population of *E. coli* followed by an increment of *Lactobacillus* counts comparatively with the control broiler group (Lei et al., 2014).

Probiotics affect the host (Fuller, 1989) and develop beneficial gut microflora that suppresses the growth of pathogens and modulate intestinal health in broiler (Shim et al., 2010; Reddy et al., 2010; Li et al., 2011; Rajput et al., 2012; Ciurescu et al., 2020).

Growth Performance

Growth performance characteristics (BW, ADG, ADFI, and FCR) are some of the most important aspects used to assess the economic benefits of broiler production (Zhang et al., 2021).

In poultry, the spore-forming probiotics affect positively FCR and are able to improve the growth and productivity of broilers in a variety of ways (Hooge et al., 2004; Jeon et al., 2014; Park et al., 2014; Ciurescu et al., 2020).

Studies from literature data have shown that the administration of *Bacillus* as probiotic product in the poultry industry can significantly

promote the growth performance of broilers (Knarreborg et al., 2008; Zhou et al., 2010; Jeong & Kim, 2014; Park & Kim, 2014; Rhayat et al., 2017). Zhang also reported that ADG was enhanced by the inclusion of *Bacillus*-based probiotics in a dose of 10^5 and 10^8 CFU/kg feed (Zhang et al., 2012, 2013). Rhayat et al. (2017) study presented that the addition of *B. subtilis* DSM 29784 in broilers significantly improved the FCR ($P < 0.05$). Reis et al. (2017) observed an improvement of FCR ($P = 0.07$) at the addition of *B. subtilis* DSM 17299 (1.6×10^9 CFU/g), results which are in agreement with other data (Jeong & Kim, 2014; Zhang et al., 2021). Bai et al. (2017) investigated the effect of *B. subtilis* fmbJ (2×10^{10} CFU/kg feed) and observed significantly improved ADG, (ADFI), and FCR of broilers ($P < 0.05$) from 21 to 42 d, respectively on the entire period. In addition, the authors noted an increase of BW in experimental groups (**BS-1**: 2519.47 ± 87.59 g; **BS-2**: 2528.10 ± 71.30 g) compared with the control (2287.34 ± 60.88 g). *B. licheniformis* DSM 28710 improved the broiler performance including BW and FCR (Trela et al., 2020). Also, *B. licheniformis* has a beneficial role on performance parameters in poultry nutrition (Lei et al., 2013; Hanuszewska et al., 2018; Musa et al., 2019). The feed supplementation with two strains of *B. subtilis* (DSM 32324 and DSM 32325) involved significantly higher BW with a lower FCR than the control group during the starter phase (Sandvang et al., 2021). Further, in the last years, spores-forming bacteria have become a topic of great interest. Single or in combination with other types of bacteria with/without the addition of minerals in the diets, probiotic-based on *Bacillus* spp. involve positive results regarding the broiler's growth promoters.

CONCLUSIONS

In summary, the present review founded on literature data shows the *Bacillus* efficacy as a probiotic and potential supplementation product for reducing antibiotics administration in the poultry industry. Based on the references from the literature, this review is focused on the influence of *Bacillus* spp. used as probiotic source in broiler feed. To point out that due to

the capacity of sporulation, the *Bacillus* group has an important advantage founded on their stability through host GIT. As an end-product, in terms of inclusion in the diets or drinking water, the use of *Bacillus* strains is easily done. Future knowledge is necessary and more investigations of the probiotic administrations in poultry should focus on *in vitro* tests for determining the action mechanism of *Bacillus*-probiotic and, as well, for clarifying the correlation between bacterial properties, level of inclusion, optimal concentration, and the host profile (age, health and production conditions, hybrid type) which can influence the purpose pursued.

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REFERENCES

- Abd El-Hack, M.E., El-Saadony, M.T., Elbestawy, A.R., El-Shall, N.A., Saad, A.M., Salem, H.M., El-Tahan, A.M., Khafaga, A.F., Taha, A.E., AbuQamar, S.F., & El-Tarabily, K.A. (2022). Necrotic enteritis in broiler chickens: disease characteristics and prevention using organic antibiotic alternatives – a comprehensive review. *Poultry Science*, 101(2), 101590.
- Abd El-Hack, M.E., El-Saadony, M.T., Shafi, M.E., Qattan, S.Y.A., Batiha, G.E., Khafaga, A.F., Abdel-Moneim, A.M. E., & Alagawany, M. (2020). Probiotic in poultry feed: A comprehensive review. *Journal of Animal Physiology and Animal Nutrition*, 104, 1835-1850.
- Abdel Baset, S., Ashour, E.A., Abd El-Hack, M.E., & El-Mekawy, M.M. (2020). Effect of different levels of pomegranate peel powder and probiotic supplementation on growth, carcass traits, blood serum metabolites, antioxidant status and meat quality of broilers. *Animal Biotechnology*, 1, 1–11.
- Abudabos, A.M., Aljumaah, M.R., Alkhulaifi, M.M., Alabdullatif, A., Suliman, G.M., & Sulaiman, R.A. (2020). Comparative effects of *Bacillus subtilis* and *Bacillus licheniformis* on live performance, blood metabolites and intestinal features in broiler inoculated with *Salmonella* infection during the finisher phase. *Microbial Pathogenesis*, 139, 103870.
- Ahmed, S.T., Islam, M.M., Mun, H.S., Sim, H.J., Kim, Y.J., & Yang, C.J. (2014). Effects of *Bacillus amyloliquefaciens* as a probiotic strain on growth performance, cecal microflora, and fecal noxious gas

- emissions of broiler chickens. *Poultry Science*, 93, 1968-1971.
- Al-Khalifa, H., Al-Nasser, A., Al-Surayee, T., Al-Kandari, S., Al-Enzi, N., Al-Sharrah, T., Ragheb, G., Al-Qalaf, & Mohammed, S. (2019). Effect of dietary probiotics and prebiotics on the performance of broiler chickens. *Journal of Poultry Sciences*, 98(10), 4465-4479.
- Amerah, A.M., Romero, L.F., Awati, A., & Ravindran, V. (2017). Effect of exogenous xylanase, amylase, and protease as single or combined activities on nutrient digestibility and growth performance of broilers fed corn/soy diets. *Poultry Science*, 96(4), 807-816.
- Apata, D.F. (2008). Growth performance, nutrient digestibility and immune response of broiler chicks fed diets supplemented with a culture of *Lactobacillus bulgaricus*. *Journal of the Science of Food and Agriculture*, 88, 1253-1258.
- Arif, M., Akteruzzaman, M., Ferdous, T.A., Islam, S.S., Das, B.C., Siddique, M.P., & Kabir, S.M. (2021). Dietary supplementation of *Bacillus*-based probiotics on the growth performance, gut morphology, intestinal microbiota and immune response. *Veterinary and Animal Science*, 1-8.
- Auger, S., Ramarao, N., Faille, C., Fouet, A., Aymerich, S., & Gohar, M. (2009). Biofilm formation and cell surface properties among pathogenic and nonpathogenic strains of the *Bacillus cereus* group. *Applied and Environmental Microbiology*, 75(20), 6616-6618.
- Bahaddad, S.A., Almalki, H.K., Alghamdi, O.A., Sohrab, S.S., Yasir, M., Azhar, E.L., & Chouayekh, H. (2022). *Bacillus* species as direct-fed microbial antibiotic alternatives for monogastric production. *Probiotics and Antimicrobial Proteins*, 1-16.
- Bai, K., Huang, Q., Zhang, J., He, J., Zhang, L., & Wang, T. (2017). Supplemental effects of probiotic *Bacillus subtilis* fmbJ on growth performance, antioxidant capacity, and meat quality of broiler chickens. *Poultry Science*, 96(1), 74-82.
- Bajagai, Y.S. (2017). Impact of *Bacillus amyloliquefaciens* probiotic strain h57 on the intestinal microbiota and broiler performance. *Doctoral thesis, School of Agriculture and Food Sciences, Australia*.
- Bajagai, Y.S., Klieve, A.V., Dart, P.J., & Bryden, W.L. (2016). Probiotics in animal nutrition: *Production, Impact and Regulation*. Rome: FAO.
- Barba-Vidal, E., Martín-Orúe, S.M., & Castillejos, L. (2019). Practical aspects of the use of probiotics in pig production: a review. *Livestock Science*, 223, 84-96.
- Barbosa, T.M., Serra, C.R., La Ragione, R.M., Woodward, M.J., & Henriques, A.O. (2005). Screening for *Bacillus* isolates in the broiler gastrointestinal tract. *Applied and Environmental Microbiology*, 71(2), 968-978.
- Barbosa, T.M., Serra, C.R., La Ragione, R.M., Woodward, M.J., & Henriques, A.O. (2011). Screening for *Bacillus* isolates in the broiler gastrointestinal tract. *Applied Environmental Microbiology*, 71, 968-978.
- Bermúdez-Humarán, L.G., Salinas, E., Ortiz, G.G., Ramírez-Jirano, L.J., Morales, J.A., & Bitzer-Quintero, O.K. (2019). From probiotics to psychobiotics: Live beneficial bacteria which act on the brain-gut axis. *Nutrients*, 11(890), 1-22.
- Bilal, M., Si, W., Barbe, F., Chevaux, E., Sienkiewicz, O., & Zhao, X. (2021). Effects of novel probiotic strains of *Bacillus pumilus* and *Bacillus subtilis* on production, gut health, and immunity of broiler chickens raised under suboptimal conditions. *Poultry Science*, 100(3), 1-11.
- Cartman, S.T., La Ragione, R.M., & Woodward, M.J. (2007). Bacterial spore formers as probiotics for poultry. *Food Science Technology Bulletin*, 4, 21-30.
- Cartman, S.T., La Ragione, R.M., & Woodward, M.J. (2008). *Bacillus subtilis* spores germinate in the chicken gastrointestinal tract. *Applied and Environmental Microbiology*, 74(16), 5254-5258.
- Cartman, S.T. & La Ragione, R.M. (2004). Spore probiotics as animal feed supplements in: *Bacterial Spore Formers: Probiotics and Emerging Applications* (Ricca, E., Henriques, A.O. & Cutting, S.M., Eds.), 155-161. Horizon Bioscience, Norfolk.
- Castañeda, C.D., Gamble, J.N., Wamsley, K.G.S., McDaniel, C.D., & Kiess, A.S. (2021). In ovo administration of *Bacillus subtilis* serotypes effect hatchability, 21-day performance, and intestinal microflora. *Poultry Science*, 100(6), 101125.
- Celandroni, F., Vecchione, A., Cara, A., Mazzantini, D., Lupetti, A., & Ghelardi, E. (2019). Identification of *Bacillus* species: Implication on the quality of probiotic formulations. *PLOS ONE*, 14(5), 1-13.
- Cervantes, H.M. (2015). Antibiotic-free poultry production: Is it sustainable? *Journal of Applied Poultry Research*, 24, 91-97.
- Chaiyawan, N., Taveeteptakul, P., Wannissorn, B., Ruengsomwong, S., Klungsunya, P., Buaban, W., & Itsaranuwat, P. (2010). Characterization and probiotic properties of *Bacillus* strains isolated from broiler. *Thai Journal of Veterinary Medicine*, 40, 207-214.
- Chauhan, A., & Singh, R. (2018). Probiotics in aquaculture: a promising emerging alternative approach. *Symbiosis*, 77(2), 99-113.
- Cheng, Y., Chen, Y., Li, X., Yang, W., Wen, C., Kang, Y., Wang, A., & Zhou, Y. (2017). Effects of synbiotic supplementation on growth performance, carcass characteristics, meat quality and muscular antioxidant capacity and mineral contents in broilers. *Journal of the Science of Food and Agriculture*, 97(11), 3699-3705. Portico.
- Ciurescu, G., Dumitru, M., & Gheorghe, A. (2021). Use of brewer's yeast (*Saccharomyces cerevisiae*) in broiler feeds to replace corn gluten meal with or without probiotic additives. *Archiva Zootechnica*, 24(1), 66-83.
- Ciurescu, G., Dumitru, M., Gheorghe, A., Untea, A.E., & Draghici, R. (2020). Effect of *Bacillus subtilis* on growth performance, bone mineralization, and bacterial population of broilers fed with different protein sources. *Poultry Science*, 99, 5960-5971.
- Clavijo, V., & Flórez, M.J.V. (2018). The gastrointestinal microbiome and its association with the control of pathogens in broiler chicken

- production: A review. *Poultry Science*, 97(3), 1006–1021.
- Cutting, S. M. (2011). *Bacillus* probiotics. *Food Microbiology*, 28, 214–220.
- Del Toro-Barbosa, M., Hurtado-Romero, A., Garcia-Amezquita, L.E., & García-Cayuela, T. (2020). Psychobiotics: Mechanisms of action, evaluation methods and effectiveness in applications with food products. *Nutrients*, 12(3896), 1-31.
- Ding, H., Zhao, X., Ma, C., Gao, Q., Yin, Y., Kong, X., & He, J. (2021). Dietary supplementation with *Bacillus subtilis* DSM 32315 alters the intestinal microbiota and metabolites in weaned piglets. *Journal of Applied Microbiology*, 130, 217–232.
- Ding, J., Dai, R., Yang, L., He, C., Xu, K., Liu, S., Zhao, W., Xiao, L., Luo, L., Zhang, Y., & Meng, H. (2017). Inheritance and Establishment of gut microbiota in chickens. *Frontiers in Microbiology*, 8, 1-11.
- Dumitru, M. & Habeanu, M. (2021). Production and evaluation of extracellular enzymes from *Bacillus licheniformis* in different raw materials used in animal feed. *Scientific Papers. Series D. Animal Science*, 64(1), 129-136.
- Dumitru, M., Habeanu, M., Lefter, N.A., & Gheorghe, A. (2020). The effect of *Bacillus licheniformis* as direct-fed microbial product on growth performance, gastrointestinal disorders and microflora population in weaning piglets. *Romanian Biotechnological Letter*, 25(6), 2060-2069.
- Dumitru, M., Habeanu, M., Sorescu, I., & Tabuc, C. (2021). Effects of *Bacillus* spp. as a supplemental probiotic in diets for weaned piglets. *South African Society for Animal Science*, 51, 578-586.
- Dumitru, M., Habeanu, M., Tabuc, C., & Jurcoane, S. (2019). Preliminary characterization of the probiotic properties of a bacterial strain for used in monogastric nutrition. *Bulletin UASVM Animal Science and Biotechnologies* 76(2), 102-108.
- Dumitru, M., Sorescu, I., Hăbeanu, M., Tabuc, C., Idriceanu, L. & Jurcoane, S. (2018). Preliminary characterisation of *Bacillus subtilis* strain use as dietary probiotic bio-additive in weaning piglet. *Journal of Food and Feed Research*, 45(2), 203-211.
- Dumitru, M., Vodnar, D.C., Elemer, S., Ciurescu, G., Habeanu, M., Sorescu, I., Georgescu, S.E., & Dudu, A. (2021). Evaluation of non-encapsulated and microencapsulated lactic acid bacteria. *Applied Sciences*, 11(9867), 1-15.
- EFSA (2019). Modification of the conditions of the authorisation of BioPlus® 2B (*Bacillus licheniformis* DSM 5749 and *Bacillus subtilis* DSM 5750) for turkeys for fattening. *EFSA Journal*, 17(6). doi.org/10.2903/j.efsa.2019.5726.
- Elisashvili, V., Kachlishvili, E., & Chikindas, M.L. (2019). Recent advances in the physiology of spore formation for *Bacillus* probiotic production. *Probiotics Antimicrobial Proteins*, 11, 731–747.
- Elshaghabe, F.M.F., Rokana, N., Gulhane, R.D., Sharma, C., & Panwar, H. (2017). *Bacillus* as potential probiotics: status, concerns, and future perspectives. *Frontiers in Microbiology*, 8, 1-15.
- European Commission (2020). Communication from the Commission to the European Parliament, the Council, the European Economic and Social Committee and the Committee of the Regions a Farm to Fork Strategy for a Fair, Healthy and Environmentally-Friendly Food System COM/2020/381 final.
- FAO & WHO (2006). Probiotics in Food. Health and Nutritional Properties and Guidelines for Evaluation. FAO Food and Nutritional, Paper No. 85.
- FAO & WHO (2021). Food and Agriculture Organization (FAO) and the World Health Organization (WHO) are to convene a Global Forum of Food Safety Regulators in Marrakech, Morocco on 28–30 January 2002, the two agencies announced today.
- Fathi, M.M., Ebeid, T.A., Al-Homidan, I., Soliman, N.K., & Abou-Emera, O.K. (2017). Influence of probiotic supplementation on immune response in broilers raised under hot climate. *British Poultry Science*, 58(5), 512-516.
- Feye, K.M., Baxter, M.F.A., Tellez-Isaias, G., Kogut, M.H., & Ricke, S.C. (2020). Influential factors on the composition of the conventionally raised broiler gastrointestinal microbiomes. *Poultry Science*, 99(2), 653–659.
- Food and Agriculture Organization/World Health Organization (2002). Guidelines for the Evaluation of Probiotics in Food: Report of a Joint FAO/WHO working group on drafting guidelines for the evaluation of probiotics in food, (London Ontario, Canada).
- Fuller, R. (1989). Probiotics in man and animals. *Journal of Applied Bacteriology*, 66, 365–378.
- Gadde, U., Oh, S. T., Lee, Y.S., Davis, E., Zimmerman, N., Rehberger, T., & Lillehoj, H. S. (2017a). The effects of direct-fed microbial supplementation, as an alternative to antibiotics, on growth performance, intestinal immune status, and epithelial barrier gene expression in broiler chickens. *Probiotics and Antimicrobial Proteins*, 9(4), 397–405.
- Gadde, U.D., Oh, S., Lee, Y., Davis, E., Zimmerman, N., Rehberger, T., et al. (2017b). Dietary *Bacillus subtilis*-based direct-fed microbials alleviate LPS induced intestinal immunological stress and improve intestinal barrier gene expression in commercial broiler chickens. *Research in Veterinary Science*, 114, 236–243.
- Gaggia, F., Mattarelli, P., & Biavati, B. (2010). Probiotics and prebiotics in animal feeding for safe food production. *International Journal of Food Microbiology*, 141, 15–28.
- Gao, Z., Wu, H., Shi, L., Zhang, X., Sheng, R., Yin, F., & Gooneratne, R. (2017). Study of *Bacillus subtilis* on growth performance, nutrition metabolism and intestinal microflora of 1 to 42 d broiler chickens. *Animal Nutrition*, 3(2), 109–113.
- Gong, J., Forster, R.J., Yu, H., Chambers, J.R., Wheatcroft, R., Sabour, P.M., & Chen, S. (2002). Molecular analysis of bacterial populations in the ileum of broiler chickens and comparison with bacteria in the cecum. *FEMS Microbiology Ecology*, 41(3), 171–179.
- Grant, A.Q., Gay, C.G., & Lillehoi, H.S. (2018). *Bacillus* spp. as direct-fed microbial antibiotic alternatives to

- enhance growth, immunity, and gut health in poultry. *Avian Pathology*, 47(4), 339-351.
- Grist, A. (2006). *Poultry Inspection: anatomy, physiology, and disease conditions*. 2nd ed., Nottingham University Press: Nottingham, UK, 262.
- Guo, M., Li, M., Zhang, C., Zhang, X., & Wu., Y. (2020). Dietary administration of the *Bacillus subtilis* enhances immune responses and disease resistance in chickens. *Frontiers in Microbiology*, 11, 1-11.
- Hanuszewska, M., Blanch, A., Kozłowski, K., & Rouault, M. (2018). Effect of *Bacillus subtilis* and *Bacillus licheniformis* inclusion in turkey diets on growth performance. *Annals of Warsaw University of Life Sciences* 57, 95–101.
- Harrington, D., Sims, M., & Kehlet, A.B. (2016). Effect of *Bacillus subtilis* supplementation in low energy diets on broiler performance. *Journal of Applied Poultry Research*, 25(1), 29–39.
- He, T., Long, S., Mahfuz, S., Wu, D., Wang X., Wei X., & Piao, X. (2019). Effects of probiotics as antibiotics substitutes on growth performance, serum biochemical parameters, intestinal morphology, and barrier function of broilers. *Animals*, 9(11), 1-10.
- Hmani, H., Daoud, L., Jlidi, M., Jalleli, K., Ben Ali, M., Hadj Brahim, A., Bargui, M., Dammak, A., & Ben Ali, M. (2017). A *Bacillus subtilis* strain as probiotic in poultry: selection based on in vitro functional properties and enzymatic potentialities. *Journal of Industrial Microbiology and Biotechnology*, 44(8), 1157–1166
- Hong, H.A., Duc, L.H., & Cutting, S.M. (2005). The use of bacterial spore formers as probiotics. *FEMS Microbiology Reviews*, 29, 813-835.
- Hong, H.A., Khaneja, R., Tam, N.M.K., Cazzato, A., Tan, S., Urdaci, M., Brisson, A., Gasbarrini, A., Barnes, I., & Cutting, S.M. (2009). *Bacillus subtilis* isolated from the human gastrointestinal tract. *Research in Microbiology*, 160(2), 134–143.
- Hooge, D.M., Ishimaru, H., & Sims, M.D. (2004). Influence of dietary *Bacillus subtilis* C-3102 spores on live performance of broiler chickens in four controlled pen Trials. *J. Appl. Poult. Res.* 13, 222–228.
- Idriceanu, L., Dumitru M., Lefter N., Gheorghe A., & Habeanu M. (2020). Dinamic plasma biochemical profile changes of weaning piglets fed diets containing different levels of *Bacillus* spp. as a probiotic product. *Archiva Zootehnică*. 23(2): 155-169.
- Jani, S.A., Chudasama, C.J., Patel, D., Bhatt, P.S., & Patel, H.N. (2012). Optimization of extracellular protease production from alkali thermo tolerant *Actinomyces: Saccharomonospora viridis* SJ-21. *Bulletin of Environment, Pharmacology and Life Sciences*, 1, 84–92.
- Jayaraman, S., Das, P.P., Saini, P.C., Roy, B., & Chatterjee, P.N. (2017). Use of *Bacillus subtilis* PB6 as a potential antibiotic growth promoter replacement in improving performance of broiler birds. *Poultry Science*, 96(8), 2614–2622.
- Jeong, J.S., & Kim, I.H. (2014). Effect of *Bacillus subtilis* C-3102 spores as a probiotic feed supplement on growth performance, noxious gas emission, and intestinal microflora in broilers. *Poultry Science*, 93, 3097–3103.
- Jiang, S., Yan, J.Y., Hu, J.J., Mohammed, A., & Cheng, H.W. (2021). *Bacillus subtilis*-Based probiotic improves skeletal health and immunity in broiler chickens exposed to heat stress. *Animals*, 11(1494), 1-21.
- Joerger, R.D. & Ganguly, A. (2017). Current status of the preharvest application of pro-and prebiotics to farm animals to enhance the microbial safety of animal products. *Microbiology Spectrum*, 5.
- Keller, D., Verbruggen, S., Cash, H., Farmer, S., & Venema, K. (2019). Spores of *Bacillus coagulans* GBI-30, 6086 show high germination, survival and enzyme activity in a dynamic, computer-controlled *in vitro* model of the gastrointestinal tract. *Beneficial Microbes*, 10(1), 77–87.
- Kerry, R.G., Pradhan, P., Samal, D., Gouda, S., Das, G., Shin H.S., & Patra, J.K. (2018). Probiotics: the ultimate nutritional supplement. *Microbial Biotechnology*. Springer, Singapore, 7, 141–152.
- Khalique, A., Zeng, D., Shoab, M., Wang, H., Qing, X., Rajput, D.S., Pan, K., & Ni, X. (2020). Probiotics mitigating subclinical necrotic enteritis (SNE) as potential alternatives to antibiotics in poultry. *AMB Express*, 10(1), 1-10.
- Khelifa, D.E., Abd El-Ghany, W.A., & Salem, H.M. (2012). Recent status of *Clostridial enteritis* affecting early-weaned rabbits in Egypt. *Life Science Journal*, 9, 2272-2279.
- Khelifa, D.G., Madian, K., El-Meneisy, A.A., Faten, F.M., & Salem H.M. (2015). Field and laboratory diagnosis of *C. perfringens* enteric infection among rabbit flocks in Egypt. *Middle East Journal of Applied Sciences*, 5, 252-261.
- Knap, I., Lund, B., Kehlet, A.B., Hofacre, C. & Mathis, G. (2010). *Bacillus licheniformis* prevents necrotic enteritis in broiler chickens. *Avian Diseases*, 54, 931–935.
- Knarreborg, A., Brockmann, E., Høybye, K., Knap, I., Lund, B., Milora, N., & Leser, T.D. (2008). *Bacillus subtilis* (DSM17299) modulates the ileal microbial communities and improves growth performance in broilers. *International Journal of Probiotics and Prebiotics*, 3, 83–88.
- Kotb, E. (2014). Purification and partial characterization of serine fibrinolytic enzyme from *Bacillus megaterium* KSK-07 isolated from kishk, a traditional Egyptian fermented food. *Applied Biochemistry and Microbiology*, 51(1), 34–43.
- La Ragione, R.M., Casula, G., Cutting, S.M. & Woodward, M. (2001). *Bacillus subtilis* spores competitively exclude *Escherichia coli* 070: K80 in poultry. *Veterinary Microbiology*, 79, 133–142.
- Latorre, J.D., Hernandez-Velasco, X., Kallapura, G., Menconi, A., Pumford, N.R., Morgan, M.J., Layton, S.L., Bielke, L.R., Hargis, B.M., & Téllez, G. (2014). Evaluation of germination, distribution, and persistence of *Bacillus subtilis* spores through the gastrointestinal tract of chickens. *Poultry Science*, 93(7), 1793–1800.
- Latorre, J.D., Hernandez-Velasco, X., Vicente, J.L., Wolfenden, R., Hargis, B.M., & Tellez, G. (2017).

- Effects of the inclusion of a *Bacillus* direct-fed microbial on performance parameters, bone quality, recovered gut microflora, and intestinal morphology in broilers consuming a grower diet containing corn distillers dried grains with solubles. *Poultry Science*, 96, 2728–2735.
- Lee, K.W., Kim, D.K., Lillehoj, H.S., Jang, S.I. & Lee, S.H. (2015). Immune modulation by *Bacillus subtilis*-based direct-fed microbials in commercial broiler chickens. *Anim. Feed. Sci. Technol.* 200, 76-85.
- Lee, K., Lillehoj, H.S., Jang, S.I., Lee, S.H., Bautista, D.A. & Siragusa, G.R. (2013). Effect of *Bacillus subtilis*-based direct-fed microbials on immune status in broiler chickens raised on fresh or used litter. *Asian-Australasian Journal of Animal Sciences*, 26, 1592–1597.
- Lee, S., Lee, J., Jin, Y., Jeong, J., Chang, Y.H., Lee, Y., Jeon, Y., & Kim, M. (2017). Probiotic characteristics of *Bacillus* strains isolated from Korean traditional soy sauce. *LWT Food Science Technology*, 79, 518–524.
- Lei, K., Li, Y.L., Yu, D.Y., Rajput, I.R., & Li, W.F. (2013). Influence of dietary inclusion of *Bacillus licheniformis* on laying performance, egg quality, antioxidant enzyme activities, and intestinal barrier function of laying hens. *Poultry Science*, 92(9), 2389–2395.
- Lei, X.J., Ru, Y.J., & Zhang, H.F. (2014). Effect of *Bacillus amyloliquefaciens*-based direct-fed microbials and antibiotic on performance, nutrient digestibility, cecal microflora, and intestinal morphology in broiler chickens. *Journal of Applied Poultry Research*, 23(3), 486–493.
- Lei, X., Piao, X., Ru, Y., Zhang, H., Péron, A., & Zhang, H. (2015). Effect of *Bacillus amyloliquefaciens*-based direct-fed microbial on performance, nutrient utilization, intestinal morphology and cecal microflora in broiler chickens. *Asian Australian Journal of Animal Science*, 28, 239–46
- Li, J., Cheng, Y., Chen, Y., Qu, H., Zhao, Y., Wen, C., & Zhou, Y. (2019). Effects of dietary synbiotic supplementation on growth performance, lipid metabolism, antioxidant status, and meat quality in Partridge shank chickens. *Journal of Applied Animal Research*, 47(1), 586–590.
- Li, W.F., Wen, J., & Hu, Z.W. (2011). Effects of *Bacillus subtilis* on growth performance, antioxidant capacity and immunity of intestinal mucosa in broilers. *Chinese Journal of Animal Science*, 47, 58–61.
- Li, Y., Zhang, H., Chen, Y.P., Yang, M.X., Zhang, L.L., Lu, Z.X., Zhou, Y.M., & Wang, T. (2015). *Bacillus amyloliquefaciens* supplementation alleviates immunological stress in lipopolysaccharide-challenged broilers at early age. *Poultry Science*, 94(7), 1504–1511.
- Liu, X., Yan, H., Lv, L., Xu, Q., Yin, C., Zhang, K., Wang, P., & Hu, J. (2012). Growth performance and meat quality of broiler chickens supplemented with *Bacillus licheniformis* in drinking water. *Asian-Australasian Journal of Animal Sciences*, 25(5), 682–689.
- Logan, N.A. & De Vos, P. (2009). In genus I. *Bacillus*; Bergey's Manual of Systematic Bacteriology. Volume 3: The Firmicutes, De Vos, P., Garrity, G., Jones, D., Krieg, N. R., Ludwig, W., Rainey, F. A., Schleifer, K. -H and Whitman, W. B. Eds., (New York: Springer), 21-127.
- Luise, D., Bosi, P., Raff, L., Amatucci, L., Virdis, S., & Trevisi, P. (2022). *Bacillus* spp. probiotic strains as a potential tool for limiting the use of antibiotics, and improving the growth and health of pigs and chickens. *Frontiers in Microbiology*, 13, 1-19.
- Ma, Y., Wang, W., Zhang, H., Wang, J., Zhang, W., Gao, J., & al. (2018). Supplemental *Bacillus subtilis* DSM 32315 manipulates intestinal structure and microbial composition in broiler chickens. *Scientific Reports*, 8(1), 33762-33768.
- Manafi, M., Khalaji, S., Hedayati, M., & Pirany, N. (2017). Efficacy of *Bacillus subtilis* and bacitracin methylene disalicylate on growth performance, digestibility, blood metabolites, immunity, and intestinal microbiota after intramuscular inoculation with *Escherichia coli* in broilers. *Poultry Science*, 96(5), 1174–1183.
- Mazanko, M.S., Gorlov, I.F., Prazdnova, E.V., Makarenko, M.S., Usatov, A.V., Bren, A.B., Chistyakov, V.A., Tutelyan, A.V., Komarova, Z.B., Mosolova, N.I., Pilipenko, D.N., Krotova, O.E., Struk, A.N., Lin, A., & Chikindas, M.L. (2017). *Bacillus* probiotic supplementations improve laying performance, egg quality, hatching of laying hens, and sperm quality of roosters. *Probiotics and Antimicrobial Proteins*, 10(2), 367–373.
- Meng, Q.W., Yan, L., Ao, X., Zhou, T.X., Wang, J.P., Lee, J.H., & Kim, I.H. (2010). Influence of probiotics in different energy and nutrient density diets on growth performance, nutrient digestibility, meat quality, and blood characteristics in growing-finishing pigs. *Journal of Animal Science*, 88, 3320–3326.
- Millette, M., Nguyen, A., Mahamad, K., & Lacroix, M. (2013). Gastrointestinal survival of bacteria in commercial probiotic products. *International Journal of Probiotics and Prebiotics*, 8(4), 149-156.
- Mingmongkolchai, S., & Panbangred, W. (2018). *Bacillus* probiotics: an alternative to antibiotics for livestock production. *Journal of Applied Microbiology*, 124, 1334-1346.
- Mountzouris, K.C., Tsirtsikos, P., Kalamara, E., Nitsch, S., Schatzmayr, G., & Fegeros, K. (2007). Evaluation of the efficacy of a probiotic containing *Lactobacillus*, *Bifidobacterium*, *Enterococcus*, and *Pediococcus* strains in promoting broiler performance and modulating cecal microflora composition and metabolic activities. *Poultry Sciences*, 86, 309–317.
- Mountzouris, K.C., Tsirtsikos, P., Palamidi, I., Arvaniti, A., Mohnl, M., Schatzmayr, G., & Fegeros, K. (2010). Effects of probiotic inclusion levels in broiler nutrition on growth performance, nutrient digestibility, plasma immunoglobulins, and cecal microflora composition. *Poultry Science*, 89, 58–67.
- Musa, B.B., Duan, Y., Khawar, H., Sun, Q., Ren, Z., Elsididg Mohamed, M.A., Abbasi, I.H.R., & Yang, X. (2019). *Bacillus subtilis* B21 and *Bacillus*

- licheniformis* B26 improve intestinal health and performance of broiler chickens with *Clostridium perfringens*-induced necrotic enteritis. *Journal of Animal Physiology and Animal Nutrition*, 103, 1039–1049.
- Mushtaq, M., Sadique, U., Said, F., Shah, M., Amanullah, H., & Anwer, F. (2021). Immunomodulatory and hepato-protective role of water based supplemented *Bacillus clausii* in broiler chicks. *Journal of the Saudi Society of Agricultural Sciences*, 21(2), 108-113.
- Neijat, M., Shirley, R.B., Barton, J., Thiery, P., Welscher, A., & Kiarie, E. (2019). Effect of dietary supplementation of *Bacillus subtilis* DSM29784 on hen performance, egg quality indices, and apparent retention of dietary components in laying hens from 19 to 48 weeks of age. *Poultry Science*, 98(11), 5622–5635.
- Nguyen, A.T.V., Nguyen, D.V., Tran, M.T., Nguyen, L.T., Nguyen, A.H., & Phan, T.N. (2015). Isolation and characterization of *Bacillus subtilis* CH16 strain from chicken gastrointestinal tracts for use as a feed supplement to promote weight gain in broilers. *Letters in Applied Microbiology*, 60(6), 580–588. Portico.
- Nguyen, D.H., Lee, K.Y., Tran, H.N., Upadhaya, S.D., Jeong, Y.J., & Kim, I.H. (2017). Influence of *Enterococcus faecium* and endo-1, 4-b-xylanase supplementation on growth performance, nutrient digestibility, fecal microflora, fecal gas emission, and meat quality in finishing pigs fed with diets based on corn–soybean meal. *Canadian Journal of Animal Science*, 98, 126–134.
- Nishiyama, T., Ashida, N., Nakagawa, K., Iwatani, S., & Yamamoto, N. (2021). Dietary *Bacillus subtilis* C-3102 supplementation enhances the exclusion of *Salmonella enterica* from chickens. *The Journal of Poultry Science*, 58(2), 138–145.
- Oakley, B.B., Lillehoj, H.S., Kogut, M.H., Kim, W.K., Maurer, J.J., Pedrosa, A., Lee, M.D., Collett, S.R., Johnson, T.J., & Cox, N.A. (2014). The chicken gastrointestinal microbiome. *FEMS Microbiology Letters*, 360(2), 100–112.
- Oladokun, S., Koehler, A., MacIsaac, J., Ibeagha-Awemu, E.M., & Adewole, D.I. (2021). *Bacillus subtilis* delivery route: effect on growth performance, intestinal morphology, cecal short-chain fatty acid concentration, and cecal microbiota in broiler chickens. *Poultry Science*, 100(3), 100809.
- Oso, A.O., Suganthi, R.U., Reddy, G.B.M., Malik, P.K., Thirumalaisamy, G., Awachat, V.B., Selvaraju, S., Arangasamy, A., & Bhatta, R. (2019). Effect of dietary supplementation with phyto-genic blend on growth performance, apparent ileal digestibility of nutrients, intestinal morphology, and cecal microflora of broiler chickens. *Poultry Science*, 98(10), 4755–4766.
- Pagnini, C., Saeed, R., Bamias, G., Arseneau, K.O., Pizarro, T.T., & Cominelli, F. (2010). Probiotics promote gut health through stimulation of epithelial innate immunity. *Proc. Natl. Acad. Sci. U.S.A.* 107, 454–459.
- Park, I., Lee, Y., Goo, D., Zimmerman, N.P., Smith, A.H., Rehberger, T., & Lillehoj, H.S. (2020). The effects of dietary *Bacillus subtilis* supplementation, as an alternative to antibiotics, on growth performance, intestinal immunity, and epithelial barrier integrity in broiler chickens infected with *Eimeria maxima*. *Poultry Science*, 99(2), 725–733.
- Park, I., Zimmerman, N.P., Smith, A.H., Rehberger, T.G., Lillehoj, Erik. P., & Lillehoj, H.S. (2020). Dietary supplementation with *Bacillus subtilis* directed microbes alters chicken intestinal metabolite levels. *Frontiers in Veterinary Science*, 7(123), 1-9.
- Park, J.H. & Kim, I.H. (2017). The effects of the supplementation of *Bacillus subtilis* RX7 and B2A strains on the performance, blood profiles, intestinal *Salmonella* concentration, noxious gas emission, organ weight and breast meat quality of broiler challenged with *Salmonella typhimurium*. *Journal of Physiology Animal Nutrition*, 99(2), 326–334.
- Park, J.H. & Kim, I.H. (2014). Supplemental effect of probiotic *Bacillus subtilis* B2A on productivity, organ weight, intestinal *Salmonella* microflora, and breast meat quality of growing broiler chicks. *Poultry Science*, 93, 2054–2059.
- Paturi, G., Phillips, M., Jones, M., & Kailasapathy, K. (2007). Immune enhancing effects of *Lactobacillus acidophilus* LAFTI L10 and *Lactobacillus paracasei* LAFTI L26 in mice. *International Journal of Food Microbiology*, 115(1), 115-118.
- Penalzo-Vazquez, A., Ma, L.M., & Rayas-Duarte, P. (2017). Isolation and characterization of *Bacillus* spp. strains as potential probiotics for poultry. *Canadian Journal of Microbiology*, 65, 762–74.
- Popov, I.V., Algburi, A., Prazdnova, E.V., Mazanko, M.S., Elisashvili, V., Bren, A.B., Chistyakov, V.A., Tkacheva, E.V., Trukhachev, V. I., Donnik, I. M., Ivanov, Y.A., Rudoy, D., Ermakov, A.M., Weeks, R.M., & Chikindas, M.L. (2021). A Review of the effects and production of spore-forming probiotics for poultry. *Animals*, 11(7), 1941.
- Rajput, I.R., Hussain, A., Li, Y.L., Zhang, X., Xu, X., Long, M.Y., et al. (2014). *Saccharomyces boulardii* and *Bacillus subtilis* B10 modulate TLRs mediated signaling to induce immunity by chicken BMDCs. *Journal of Cellular Biochemistry*, 115, 189–198.
- Rajput, I.R., Li, L.Y., Xin, X., Wu, B.B., Juan, Z.L., Cui, Z.W., Yu, D.Y., & Li, W.F. (2013). Effect of *Saccharomyces boulardii* and *Bacillus subtilis* B10 on intestinal ultrastructure modulation and mucosal immunity development mechanism in broiler chickens. *Poultry Science*, 92(4), 956–965.
- Ramlucken, U., Laloo, R., Roets, Y., Moonsamy, G., Van Rensburg, C.J., & Thantsha, M.S. (2020a). Advantages of *Bacillus*-based probiotics in poultry production. *Livestock Science*, 241, 104215.
- Ramlucken, U., Roets, Y., Ramchuran, S.O., Moonsamy, G., Rensburg, C.J., Thantsha, M.S., & Laloo, R. (2020b). Isolation, selection and evaluation of *Bacillus* spp. as potential multi-mode probiotics for poultry. *Journal of General and Applied Microbiology*, 3-30.
- Ramlucken, U, Ramchuran, S.O., Moonsamy, G., Laloo, R., Thantsha, M.S., & Van Rensburg, C.J. (2020c). A

- novel *Bacillus* based multi-strain probiotic improves growth performance and intestinal properties of *Clostridium perfringens* challenged broilers. *Poultry Science*, 1, 331–342.
- Ran, T., Gomaa, W.M.S., Shen, Y.Z., Saleem, A.M., Yang, W.Z., & McAllister, T.A. (2019). Use of naturally sourced feed additives (*Lactobacillus* fermentation products and enzymes) in growing and finishing steers: Effects on performance, carcass characteristics and blood metabolites. *Animal Feed Science and Technology*, 254, 114190.
- Ravindran, V. (2013). Feed enzymes: The science, practice, and metabolic realities. *J. Appl. Poult. Res.*, 22, 628–636.
- Reddy, P.V.M., Kondal, R.K., Kuhad, R.C., Shashi, K.M., & Gnana, P.M. (2010). Effect of supplementation of enzymes and probiotics on performance of broiler chicken. *Indian Journal of Poultry Science*, 45, 361–363.
- Reis, M.P., Fassani, E.J., Júnior, A.A.P.G., Rodrigues, P.B., Bertechini, A.G., Barrett, N., Persia, M.E., & Schmidt, C.J. (2017). Effect of *Bacillus subtilis* (DSM 17299) on performance, digestibility, intestine morphology, and pH in broiler chickens. *Journal of Applied Poultry Research*, 26(4), 573–583.
- Rhayat, L., Jacquier, V., Brinch, K.S., Nielsen, P., Nelson, A., Geraert, P.A., & Devillard, E. (2017). *Bacillus subtilis* strain specificity affects performance improvement in broilers. *Poultry Science*, 96(7), 2274–2280.
- Salem, H.M., & Attia, M.M. (2021). Accidental intestinal myiasis caused by *Musca domestica* L. (Diptera: Muscidae) larvae in broiler chickens: a field study. *International Journal of Tropical Insect Science*, 41(4), 2549–2554.
- Samanya, M. & Yamauchi, K.E. (2002). Histological alterations of intestinal villi in chickens fed dried *Bacillus subtilis* var. natto. *Comparative Biochemistry and Physiology Part A: Molecular Integrative Physiology*, 133, 95–104.
- Sandvang, D., Skjot-Rasmussen, L., Cantor, M.D., Mathis, G.F., Lumpkins, B.S., & Blanch, A. (2021). Effects of feed supplementation with 3 different probiotic *Bacillus* strains and their combination on the performance of broiler chickens challenged with *Clostridium perfringens*. *Poultry Science*, 100(4), 100982.
- Scanes, C.G. (2014). *Sturkie's Avian Physiology*, 6th ed.; Elsevier: Amsterdam, The Netherlands, 1056.
- Schrezenmeir, J. & de Vrese, M. (2001). Probiotics, prebiotics, and synbiotics—approaching a definition. *The American Journal of Clinical Nutrition*, 73(2), 361s–364s.
- Sen, S., Ingale, S.L., Kim, Y.W., Kim, J.S., Kim, K.H., Lohakare, J.D., Kim, E.K., Kim, H.S., Ryu, M.H., Kwon, I.K., & Chae, B.J. (2012). Effect of supplementation of *Bacillus subtilis* LS 1-2 to broiler diets on growth performance, nutrient retention, caecal microbiology and small intestinal morphology. *Research in Veterinary Science*, 93(1), 264–268.
- Shah, K.R., & Bhatt, S.A. (2011). Purification and characterisation of lipase from *Bacillus subtilis* Pa2. *Journal of Biochemical Technology*, 3, 292–295.
- Shim, Y.H., Shinde, P.L., Choi, J.Y., Kim, J.S., Seo, D.K., Pak, J.I., Chae, B.J., & Kwon, I.K. (2010). Evaluation of multi-microbial probiotics produced by submerged liquid and solid substrate fermentation methods in broilers. *Asian-Australasian Journal of Animal Science* 23, 521–529.
- Shivaramaiah, S., Pumford, N.R., Morgan, M.J., Wolfenden, R.E., Wolfenden, A.D., Torres-Rodríguez, A., Hargis, B.M., & Téllez, G. (2011). Evaluation of *Bacillus* species as potential candidates for direct-fed microbials in commercial poultry. *Poultry Science*, 90(7), 1574–1580.
- Sikandar, A., Zaneb, H., Nasir, A., Adil, M., Ali, H.M., Muhammad, N., Rehman, T., Rehman, A., & Rehman, H.F. (2021). Effects of *Bacillus subtilis* on performance, immune system and gut in *Salmonella*-challenged broilers. *South African Journal of Animal Science*, 50(5), 1-9.
- Simon, O., Jadamus, A., & Vahjen, W. (2001). Probiotic feed additives—effectiveness and expected modes of action. *Journal of Animal Science*, 10, 51–67.
- Sinol, S., Ingale, S.L., Kim, Y.W., Kim, J.S., Kima, K.H., Lohakarea, J.D., Kim, E.K., Kim, H.S., Ryu, M.H., Kwon, I.K., & Chae, B.J. (2012). Effect of supplementation of *Bacillus subtilis* LS 1–2 to broiler diets on growth performance, nutrient retention, caecal microbiology and small intestinal morphology. *Research in Veterinary Science*, 93, 264–268. UK.
- Sobczak, A. & Kozłowski, K. (2015). The effect of a probiotic preparation containing *Bacillus subtilis* ATCC PTA-6737 on egg production and physiological parameters of laying hens. *Annals of Animal Sciences*, 15(3), 711–723.
- Sorescu, I., Dumitru, M., & Ciurescu, G. (2019). *Lactobacillus* spp. and *Enterococcus faecium* strains isolation, identification, preservation and quantitative determinations from turkey gut content. *Romanian Biotechnological Letter*, 24(1), 41–49.
- Sosa, N., Gerbino, E., Golowczyk, M.A., Schebor, C., Gómez-Zavaglia, A., Tymczyszyn, E.E. (2016). Effect of galacto-oligosaccharides: maltodextrin matrices on the recovery of *Lactobacillus plantarum* after spray-drying. *Frontiers in Microbiology*, 3(7), 2-8.
- Sumi, C.D., Yang, B.W., Yeo, I.C., & Hahm, Y.T. (2015). Antimicrobial peptides of the genus *Bacillus*: a new era for antibiotics. *Canadian Journal of Microbiology*, 61(2), 93–103. doi.org/10.1139/cjm-2014-0613.
- Tang, R.Y., Wu, Z.L., Wang, G.Z., & Liu, W.C. (2017). The effect of *Bacillus amyloliquefaciens* on productive performance of laying hens. *Italian Journal of Animal Science*, 17(2), 436–441.
- Tarradas, J., Tous, N. & Esteve-Garcia, E. (2020). The control of intestinal inflammation: A major objective in the research of probiotic strains as alternatives to antibiotic growth promoters in poultry. *Microorganisms* 8(2), 148.
- Tazehabadi, M.H., Algburi, A., Popov, I.V., Ermakov, A.M., Chistyakov, V.A., Prazdnova, E.V., Weeks, R., & Chikindas, M.L. (2021). Probiotic Bacilli inhibit *Salmonella* biofilm formation without killing planktonic cells. *Frontiers in Microbiology*, 1-12.

- Teo, A.Y. & Tan, H.M. (2007). Evaluation of the performance and intestinal gut microflora of broilers fed on corn-soy diets supplemented with *Bacillus subtilis* PB6 (CloSTAT). *Journal of Applied Poultry Research*, 16, 296–303.
- Tidjani Alou, M., Rathored, J., Khelaifia, S., Michelle, C., Brah, S., Diallo, B.A., Raoult, D., & Lagier, J.C. (2015). *Bacillus rubiinfantis* sp. nov. strain mt2T, a new bacterial species isolated from human gut. *New Microbes and New Infections*, 8, 51–60.
- Trela, J., Kierończyk, B., Hautekiet, V., & Józefiak, D. (2020). Combination of *Bacillus licheniformis* and salinomycin: effect on the growth performance and gut microbial populations of broiler chickens. *Animals*, 10(5), 889.
- Uraisha, R., Ramchuran, S.O., Moonsamy, G., Van Rensburg, C. J., Thantsha, M.S., & Lallo, R. (2020). Production and stability of a multi-strain *Bacillus* based probiotic product for commercial use in poultry. *Biotechnology Reports*, 29, 1-8.
- Van der Aar, P.J., Molist, F., & Van der Klis, J.D. (2017). The central role of intestinal health on the effect of feed additives on feed intake in swine and poultry. *Animal Feed Science and Technology*, 233, 64–75.
- Wang, X., Peebles, E.D., Kiess, A.S., Wamsley, K.G.S., Zhai, W. (2019). Effects of coccidial vaccination and dietary antimicrobial alternatives on the growth performance, internal organ development, and intestinal morphology of *Eimeria*-challenged male broilers. *Poultry Science*, 98, 2054–2065.
- Wang, Y., Zhang, H., Zhang, L., Liu, W., Zhang, Y., Zhang, X., & Sun, T. (2010). *In vitro* assessment of probiotic properties of *Bacillus* isolated from naturally fermented congee from Inner Mongolia of China. *World Journal of Microbiology and Biotechnology*, 26(8), 1369–1377.
- Xu, X., Huang, Q., Mao, Y., Cui, Z., Li, Y., Huang, Y., Rajput, I.R., Yu, D., & Li, W. (2012). Immunomodulatory effects of *Bacillus subtilis* (natto) B4 spores on murine macrophages. *Microbiology and Immunology*, 56(12), 817–824.
- Zaghari, M., Zahroojian, N., Riahi, M., & Parhizkar, S. (2015). Effect of *Bacillus subtilis* spore (GalliPro®) nutrients equivalency value on broiler chicken performance. *Italian Journal of Animal Science*, 14(1), 3555, 94-98.
- Zhang, L., Cao, G.T., Zeng, X.F., Zhou, L., Ferket, P.R., Xiao, Y.P., Chen, A.G., & Yang, C.M. (2014). Effects of *Clostridium butyricum* on growth performance, immune function, and cecal microflora in broiler chickens challenged with *Escherichia coli* K88. *Poultry Science*, 93(1), 46–53.
- Zhang, S., Zhong, G., Shao, D., Wang, Q., Hu, Y., Wu, T., Ji, C., & Shi, S. (2021). Dietary supplementation with *Bacillus subtilis* promotes growth performance of broilers by altering the dominant microbial community. *Poultry Science*, 100(3), 1-13.
- Zhang, W., Zhu, Y.H., Zhou, D., Wu, Q., Song, D., Dicksved, J., & Wang, J.F. (2017). Oral administration of a select mixture of *Bacillus* probiotics affects the gut microbiota and goblet cell function following *Escherichia coli* challenge in newly weaned pigs of genotype muc4 that are supposed to be enterotoxigenic *E. coli* F4ab/ac receptor negative. *Journal Applied and Environmental Microbiology*, 83(3), 1-18.
- Zhang, Z.F., Cho, J.H., & Kim I.H. (2013). Effects of *Bacillus subtilis*, ubt-mo 2, on growth performance, relative immune organ weight, gas concentration in excreta, and intestinal microbial shedding in broiler chickens. *Livestock Science*, 155, 343–347.
- Zhang, Z.F., Zhou, T.X., Ao, X., & Kim, I.H. (2012). Effects of β glucan and *Bacillus subtilis*, on growth performance, blood profiles, relative organ weight and meat quality in broilers fed maize–soybean meal based diets. *Livestock Sciences*, 150, 419–424.
- Zhen, W., Shao, Y., Gong, X., Wu, Y., Geng, Y., Wang, Z., & Guo, Y. (2018). Effect of dietary *Bacillus coagulans* supplementation on growth performance and immune responses of broiler chickens challenged by *Salmonella enteritidis*. *Poultry Science*, 97(8), 2654–2666.
- Zhou, X., Wang, Y., Gu, Q., & Li, W. (2010). Effect of dietary probiotic, *Bacillus coagulans*, on growth performance, chemical composition, and meat quality of Guangxi Yellow chicken. *Poultry Science*, 89(3), 588–593.